



Development Support Document
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Ethylene Dichloride

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TEXAS COMMISSION ON ENVIRONMENTAL QUALITY

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Acronyms and Abbreviations

Acronyms and Abbreviations	Definitions
ACGIH	American Conference of Industrial Hygienists
ADAF	age-dependent adjustment factor
AEGL	Acute Exposure Guideline Level
ALT	alanine aminotransferase
AMCV	Air Monitoring Comparison Value
ATSDR	Agency for Toxic Substances and Disease Registry
BMC	benchmark concentration
BMR	benchmark response
°C	degrees Celsius
CNS	central nervous system
d	day
DSD	development support document
ESL	Effects Screening Level
^{acute} ESL	acute health-based Effects Screening Level for chemicals meeting minimum database requirements
^{acute} ESL _{generic}	acute health-based Effects Screening Level for chemicals not meeting minimum database requirements
^{acute} ESL _{odor}	acute odor-based Effects Screening Level
^{acute} ESL _{veg}	acute vegetation-based Effects Screening Level
^{chronic} ESL _{nonthreshold(c)}	chronic health-based Effects Screening Level for nonthreshold dose response cancer effect
^{chronic} ESL _{nonthreshold(nc)}	chronic health-based Effects Screening Level for nonthreshold dose response noncancer effects
^{chronic} ESL _{threshold(c)}	chronic health-based Effects Screening Level for threshold dose response cancer effects
^{chronic} ESL _{threshold(nc)}	chronic health-based Effects Screening Level for threshold dose response noncancer effects

Acronyms and Abbreviations	Definitions
chronic ^{ESL_{veg}}	chronic vegetation-based Effects Screening Level
ET	extrathoracic
GD	gestational day
GLP	good laboratory practice
h	hour(s)
H _{b/g}	blood:gas partition coefficient
(H _{b/g}) _A	blood:gas partition coefficient, animal
(H _{b/g}) _H	blood:gas partition coefficient, human
Hg	mercury
HEC	human equivalent concentration
HPV	high production volume
HQ	hazard quotient
i.p.	intraperitoneal
IARC	International Agency for Research on Cancer
kg	kilogram
LOAEL	lowest observed adverse effect level
LOEL	lowest observed effect level
MW	molecular weight
µg	microgram(s)
µg/m ³	micrograms per cubic meter
mg	milligram(s)
mg/L	milligrams per liter
mg/m ³	milligrams per cubic meter
min	minute(s)
MOA	mode of action
n	number

Acronyms and Abbreviations	Definitions
N/A	Not applicable
NCI	National Cancer Institute
NOAEL	no observed adverse effect level
NOEL	no observed effect level
NTP	National Toxicology Program
OEHHA	Office of Environmental Health Hazard Assessment
PND	postnatal day
POD	point of departure
POD _{ADJ}	point of departure adjusted for exposure duration
POD _{HEC}	point of departure adjusted for human equivalent concentration
ppb	parts per billion
ppm	parts per million
ReV	reference value
RGDR	regional gas dose ratio
SD	Sprague-Dawley
TAMIS	Texas Air Monitoring Information System
TCEQ	Texas Commission on Environmental Quality
TD	Toxicology Division
UF	uncertainty factor
UF _H	interindividual or intraspecies human uncertainty factor
UF _A	animal to human uncertainty factor
UF _{Sub}	subchronic to chronic exposure uncertainty factor
UF _L	LOAEL to NOAEL uncertainty factor
UF _D	incomplete database uncertainty factor
URF	unit risk factor
USEPA	United States Environmental Protection Agency

Acronyms and Abbreviations	Definitions
wk	week(s)
yr	year(s)

1 **Chapter 1 Summary Tables**

2 Table 1 for air monitoring and Table 2 for air permitting provide a summary of health- and
3 welfare-based values resulting from an acute and chronic evaluation of ethylene dichloride (1,2-
4 dichloroethane; EDC). Please refer to Section 1.6.2 of the TCEQ Toxicity Factor Guidelines
5 (2012) for an explanation of values used for review of ambient air monitoring data and air
6 permitting. Table 3 provides summary information on EDC’s physical/chemical properties.

7 **Table 1. Air Monitoring Comparison Values (AMCVs) for Ambient Air**

Short-Term Values	Concentration	Notes
Acute ReV [1 h]	2,200 µg/m ³ (550 ppb) 1-h Short-Term Health	Critical Effect(s): Very slight degeneration and necrosis of the olfactory epithelium in rats
Acute ReV [24 h]	380 µg/m ³ (93 ppb) 24-h Short-Term Health	Critical Effect(s): Very slight degeneration and necrosis of the olfactory epithelium in rats
^{acute} ESL _{odor}	---	Pleasant odor; odor-based ESL significantly above health-based values
^{acute} ESL _{veg}	---	No data on vegetation effects found
Long-Term Values	Concentration	Notes
Chronic ReV	44 µg/m ³ (11 ppb)	Critical Effect(s): Suggestive liver and kidney toxicity in rats
^{chronic} ESL _{nonthreshold(c)}	3.7 µg/m ³ (0.91 ppb) Long-Term Health	Critical Effect(s): Increased incidence of liver hemangiosarcomas in mice
^{chronic} ESL _{veg}	---	No data on vegetation effects found

8 Abbreviations for Tables 1 and 2: **ppb**, parts per billion; **µg/m³**, micrograms per cubic meter; **h**,
9 hour; **ESL**, Effects Screening Level; **AMCV**, Air Monitoring Comparison Value; **HQ**, hazard
10 quotient; **ReV**, Reference Value; ^{acute}**ESL**, acute health-based ESL; ^{acute}**ESL_{odor}**, acute odor-
11 based ESL; ^{acute}**ESL_{veg}**, acute vegetation-based ESL; ^{chronic}**ESL_{nonthreshold(c)}**, chronic health-based
12 ESL for nonthreshold dose-response cancer effect; ^{chronic}**ESL_{threshold(nc)}**, chronic health-based
13 ESL for threshold dose-response noncancer effects; and ^{chronic}**ESL_{veg}**, chronic vegetation-based
14 ESL

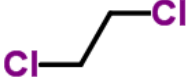
1 **Table 2. Air Permitting Effects Screening Levels (ESLs)**

Short-Term Values	Concentration	Notes
^{acute} ESL [1 h] (HQ = 0.3)	650 µg/m ³ (160 ppb) ^a Short-Term ESL for Air Permit Reviews	Critical Effect: Very slight degeneration and necrosis of the olfactory epithelium in rats
^{acute} ESL _{odor}	---	Pleasant odor; odor-based ESL significantly above health-based values
^{acute} ESL _{veg}	---	No data on vegetation effects found
Long-Term Values	Concentration	Notes
^{chronic} ESL _{threshold(nc)} (HQ = 0.3)	13 µg/m ³ (3.3 ppb) ^b	Critical Effect: Suggestive liver and kidney toxicity in rats
^{chronic} ESL _{nonthreshold(c)}	3.7 µg/m ³ (0.91 ppb) Long-Term ESL for Air Permit Reviews	Critical Effect(s): Increased incidence of liver hemangiosarcomas in mice
^{chronic} ESL _{veg}	---	No data on vegetation effects found

2 ^a Based on the acute ReV of 2,200 µg/m³ (550 ppb) multiplied by 0.3 and rounded to two
3 significant digits to account for cumulative and aggregate risk during the air permit review.

4 ^b Based on the chronic ReV of 44 µg/m³ (11 ppb) multiplied by 0.3 and rounded to two
5 significant digits to account for cumulative and aggregate risk during the air permit review.

1 **Table 3. Chemical and Physical Data**

Parameter	Value	Reference
Molecular Formula	C ₂ H ₄ Cl ₂	ATSDR 2001
Chemical Structure		ChemSpider 2014
Molecular Weight	98.96 g/mol	ATSDR 2001
Physical State at 25°C	Heavy liquid	ATSDR 2001
Color	Colorless	ATSDR 2001
Odor	Pleasant	ATSDR 2001
CAS Registry Number	107-06-2	ATSDR 2001
Synonyms	Ethylene dichloride; 1,2-dichloroethane; EDC; Dutch liquid	ATSDR 2001
Solubility in water	8.69 x 10 ³ mg/L at 20°C	ATSDR 2001
Log K _{ow}	1.48	ATSDR 2001
Vapor Pressure	79.1 mm Hg at 25°C	ATSDR 2001
Relative Vapor Density (air = 1)	1.23 g/cm ³ at 25°C	ATSDR 2001
Melting Point	-35.5°C	ATSDR 2001
Boiling Point	83.5°C	ATSDR 2001
Conversion Factors	1 µg/m ³ = 0.247 ppb 1 ppb = 4.05 µg/m ³ at 25°C	ATSDR 2001

2 **Chapter 2 Major Sources and Uses**

3 EDC is a manufactured chemical that is not found naturally in the environment. EDC is primarily
4 used in the production of vinyl chloride, which is used to manufacture many other products such
5 as plastic and polyvinyl chloride (PVC) products, construction materials, furniture and
6 automobile upholstery, wall coverings, housewares, packaging materials, and automobile parts
7 (ATSDR 2001). In the past, EDC was used as a gasoline additive to scavenge inorganic lead
8 compounds, but the transition to unleaded gasoline removed the need for this addition (OEHHA
9 2000).

1 EDC is mostly released into the environment through the air, although it can also be released into
2 soil and water. Although large amounts of EDC from a leak or spill may remain in the soil for
3 more than 40 days, small amounts typically evaporate very quickly and are mostly found in air.
4 Once in the air, EDC is broken down in a slow process by sunlight and has a half-life of
5 approximated 73 days (ATSDR 2001). The Agency for Toxic Substances and Disease Registry
6 (ATSDR) estimates that daily atmospheric concentrations range from 0.01-0.1 ppb for most
7 rural, suburban, and urban areas, although concentrations may be higher near point sources. In
8 2014, annual averages for EDC from 50 canister samplers across the state of Texas ranged from
9 not detected to 0.18 ppb (Texas Air Monitoring Information System, TAMIS). The four industry-
10 sponsored monitors located at the Formosa Plant, a site with historical EDC emissions, had 2014
11 annual averages ranging from 0.27 ppb to 0.52 ppb.

12 **Chapter 3 Acute Evaluation**

13 The Development Support Document (DSD) is a summary of the key and supporting studies and
14 procedures used by the Toxicology Division (TD) to derive inhalation toxicity values. This
15 section is based on a review of current literature as well as background readings in ATSDR
16 (2001) and USEPA (2010), which describe in detail the acute toxicity of EDC.

17 **3.1 Health-Based Acute 1-Hour ReV and ^{acute}ESL**

18 **3.1.1 Physical/Chemical Properties**

19 EDC is produced as a liquid. It is soluble in water and other organic solvents such as alcohols
20 and ethers, and volatilization occurs easily as well. The primary physical and chemical properties
21 of EDC are summarized in Table 3.

22 **3.1.2 Key and Supporting Studies**

23 **3.1.2.1 Human Studies**

24 Several studies have described the effects of acute or occupational exposure to high levels of
25 EDC. Although significant detail about the exposure parameters is typically lacking, such as air
26 concentrations and exact times of exposure, the studies all seem to point to similar biological
27 targets for EDC. Disruptions in the whole body processes such as the central nervous system
28 (CNS), and damage to multiple organs including the liver, kidney and lungs suggests that
29 although these short term studies may be lacking in exposure information, EDC appears to have
30 both a point-of-entry (POE) and systemic mode of action. The following studies were found as
31 individual research papers and/or detailed in ATSDR (2001):

- 32 • Nouchi et al. (1984) reported a case study of a 51-year-old man who was exposed to a “thick
33 vapor” of EDC for about 30 minutes (min) while cleaning out a tank. Specific information
34 was not given on the air concentration of EDC, the possibility of other contaminants in the
35 tank, or the precise route of exposure, although the report assumed that it was primarily

1 through inhalation with some dermal exposure. Coworkers found the man squatting and
2 lethargic in the tank, but he became more alert shortly after being removed. He vomited
3 sporadically overnight and was found drowsy and in respiratory distress approximately 20
4 hours after the initial exposure. He was admitted to the hospital “delirious and tremulous”
5 and fell into a coma shortly after. He died five days later from cardiac arrhythmia. An
6 autopsy revealed congestion in the lungs, degeneration in the myocardium, liver and renal
7 tubular necrosis, and shrunken nerve cells in the brain.

- 8 • Cheng et al. (1999) examined liver function in workers exposed to EDC and vinyl chloride
9 monomer (VCM). EDC is used in the manufacturing of VCM, so occupational exposures
10 often occur concurrently. Cheng et al. surveyed 251 workers from four different VCM plants
11 (mean age 39 years; mean duration of employment 1.1 years) and conducted personal and
12 area sampling. Blood tests for markers of liver function including serum aspartate
13 aminotransferase (AST), alanine aminotransferase (ALT), and gamma-glutamyltransferase
14 (GGT) were performed on each of the subjects. Area sampling showed median EDC
15 concentrations generally below 0.5 ppm, with some higher concentrations found in specific
16 area of the plants (1.31 ppm in the oxychlorination area and 2.63 ppm in the EDC loading
17 area). Personal samplers were consistent with area monitors, with EDC unloading operators
18 having a median exposure of 0.77 ppm EDC (range of 0.17 – 333.70 ppm). The subjects
19 were placed into three categories based on their job/exposure: low-EDC-low-VCM,
20 moderate-EDC-low-VCM, and low-EDC-moderate-VCM. When using the low-EDC-low-
21 VCM group as a reference, both the moderate-EDC-low-VCM and low-EDC-moderate-
22 VCM groups had higher odds ratio (OR) for developing an abnormal ALT. The moderate-
23 EDC-low-VCM group also had a higher OR for developing an abnormal AST. GGT was not
24 associated with exposure to EDC or VCM. These results suggest that co-exposure to EDC
25 and VCM may lead to decreased liver function; however the effects of EDC exposure alone
26 were not examined.
- 27 • Bowler et al. (2003) reported on the effects of EDC exposure in a group of 221 hazardous
28 waste clean-up workers. This study was done as part of a lawsuit, however the article was
29 peer reviewed and published in a scientific journal. A large spill occurred in the southern US
30 where over 69 million pounds of EDC leaked from an underground pipeline. Approximately
31 1600 hazardous waste clean-up workers were hired to clean up the spill, which involved
32 standing in contaminated water and soil without protective equipment. Although dermal
33 exposure was thought to be the primary route of exposure, inhalation of EDC also likely
34 contributed to their exposure. No estimates were given on the exposure concentrations or
35 durations. Approximately 800 of the workers were examined by physicians, with about 400
36 having mental health and neurological complaints. Of these 400 workers, 221 were referred
37 by the physicians for this study and a complete neuropsychological evaluation. It is important
38 to note that these workers were referred for the study because they had what the physicians
39 considered to be the highest exposures and the most health complaints, so this study has an
40 inherent bias. After several rounds of exclusions, the researchers narrowed the pool down to
41 137 workers. The researchers concluded that the “available hazardous waste clean-up
42 workers had significantly poorer performance on the neuropsychological tests and

1 significantly more emotional dysfunction compared to normative test data”. The lack of
2 exposure data and the bias inherent in this study make it difficult to use in the development
3 of a ReV.

4 A lack of well-conducted human studies has led to the use of an animal study to derive the 1-
5 hour (h) acute ReV and ESL.

6 **3.1.2.2 Animal Studies**

7 **3.1.2.2.1 Key Animal Study (Hotchkiss et al. 2010)**

8 Hotchkiss et al. (2010) conducted an acute toxicological and neurological evaluation of the
9 effects of EDC inhalation in rats. These studies were in accordance with good laboratory practice
10 (GLP) regulations and followed regulatory guidance set by the USEPA for testing of high
11 production value (HPV) chemicals. Equal numbers of male and female Fischer 344 rats were
12 exposed to target concentrations of 0, 200, 600, and 2000 ppm EDC for 4 h. Because significant
13 histological changes were observed in the lowest exposure group, an additional set of animals
14 was exposed to concentrations of 0, 50, 100, and 150 ppm for 8 h and 50 ppm for 4 h. Animals
15 were divided into three groups based on their exposure and histological examination:

- 16 • Acute inhalation toxicity group 1: equal numbers of male and female Fischer 344 rats
17 with target concentrations of 0, 200, 600, and 2000 ppm (mean analytical concentrations
18 0, 196.4, 607.8, and 2029.0) for 4 h (5/sex/group)
- 19 • Acute inhalation toxicity group 2: equal numbers of male and female Fischer 344 rats
20 with target concentrations of 0, 50, 100, and 150 ppm (mean analytical concentrations 0,
21 52.8, 107.5, 155.8 ppm) for 8 h and 50 ppm (mean analytical concentration 52.4) for 4 h
22 (5/sex/group)
- 23 • Acute neurotoxicity group: equal numbers of male and female Fischer 344 rats with
24 target concentrations of 0, 200, 600, and 2000 ppm (mean analytical concentrations 0,
25 196.4, 607.8, and 2029.0 ppm) for 4 h (10/sex/group)

26 All rats were exposed in 2 m³ or 4 m³ stainless steel and glass whole-body exposure chambers.
27 EDC was vaporized in atmospheric nitrogen using a glass J-tube method, then mixed in the
28 exposure chambers until the test concentrations were achieved. EDC concentrations were
29 measured every 2 h, and the mean concentrations were calculated for each exposure group.
30 Animals were weighed and clinically evaluated throughout the exposure duration, and were
31 sacrificed 24 h after the last exposure. Body weights, gross organ weights and pathology, and
32 detailed histopathology were examined following exposure.

33 There were no treatment-related clinical abnormalities observed in any of the animals tested at
34 the end of the exposure period. Rats at the highest exposure concentration (2000 ppm) had an
35 incoordinated gait that the author’s attributed to EDC exposure that resolved itself after day 2.

1 Body weights were decreased in the two highest exposure groups (600 and 2000 ppm) by an
2 average of 6-11 grams (g) (3-5%), which was more than the weight loss seen in the 200 ppm
3 exposure group and controls (3-4 g, 1.5-2% loss). None of these differences were statistically
4 significant but they were dose-dependent. No changes in body weight were observed at
5 concentrations of 150 ppm or less. There were no gross pathological observations on the organs
6 that were treatment related. Rats exposed to the highest concentration of 2000 ppm had
7 statistically significant increases in the relative and absolute adrenal gland weights. Male rats
8 also had slightly increased liver weights, but the female rats had slightly decreased liver weights,
9 so it was not determined whether this change was treatment related. No other gross organ
10 changes were observed.

11 Histopathological effects were observed in the nasal tissue, kidneys, and liver of EDC exposed
12 rats. The incidence of nasal lesions in male and female rats is presented in Table 4 and Table 5,
13 respectively. Very slight to slight degeneration and necrosis of the olfactory epithelium occurred
14 in all of the rats exposed to 600 and 2000 ppm, and 3 out of 5 males and 4 out of 5 males in the
15 200 ppm group. Because these nasal effects were seen in all of the experimental groups, a second
16 cohort was exposed to concentrations of 50, 100, and 150 ppm for 8 h. Degeneration and
17 necrosis of the olfactory epithelium was also observed in 4 out of 5 males and 5 out of 5 females
18 in the 150 ppm group, and 1 out of 5 males and 3 out of 5 females in the 100 ppm group.
19 Exposure to 50 ppm gave results similar to those seen in the control animals. An additional
20 group of rats sacrificed 15 d after the last exposure showed no degeneration in the nasal
21 epithelium, but signs of regeneration were observed in some, suggesting that these effects may
22 be reversible. A statistical analysis was not conducted. The no observed effect level (NOEL) for
23 degeneration and necrosis of the olfactory epithelium is 50 ppm.

24 **Table 4. Incidence of olfactory epithelium degeneration and necrosis in male rats.**

EDC (ppm)	n per group	Very Slight, Unilateral	Very Slight, Bilateral	Slight	Moderate	Total
0	5 ^a /5 ^b /5 ^c	0/0/0	0/0/0	0/0/0	0/0/0	0/0/0
50	5 ^a /5 ^b	0/0	0/0	0/0	0/0	0/0
100	5 ^b	1	0	0	0	1
150	5 ^b	2	2	0	0	4
200	5 ^a /5 ^c	1/0	2/0	0/0	0/0	3/0
600	5 ^a /5 ^c	0/0	1/0	4/0	0/0	5/0
2000	5 ^a /5 ^c	0/0	1/0	4/0	0/0	5/0

25 ^a 4-h exposure, 24 h sacrifice

26 ^b 8-h exposure, 24 h sacrifice

27 ^c 4-h exposure, 15 d sacrifice

1 **Table 5. Incidence of olfactory epithelium degeneration and necrosis in female rats.**

EDC (ppm)	n per group	Very Slight, Unilateral	Very Slight, Bilateral	Slight	Moderate	Total
0	5 ^a /5 ^b /5 ^c	0/0/0	0/0/0	0/0/0	0/0/0	0/0/0
50	5 ^a /5 ^b	0/0	0/0	0/0	0/0	0/0
100	5 ^b	3	0	0	0	3
150	5 ^b	0	4	1	0	5
200	5 ^a /5 ^c	2/0	1/0	1/0	0/0	4/0
600	5 ^a /5 ^c	0/0	1/0	4/0	0/0	5/0
2000	5 ^a /5 ^c	0/0	2/0	2/0	1/0	5/0

2 ^a 4-h exposure, 24 h sacrifice

3 ^b 8-h exposure, 24 h sacrifice

4 ^c 4-h exposure, 15 d sacrifice

5 At 2000 ppm, male rats had very slightly increased basophilia in the renal tubular epithelium and
6 females had degeneration and individual cell necrosis of a segmented portion of the nephron.
7 Kidney effects were not observed in any other exposure group. Male rats exposed to 2000 ppm
8 had very slight macrophage aggregates and less hepatocyte cytoplasmic vacuolation in the liver
9 compared to controls. Female rats had altered cytoplasmic homogeneity of hepatocytes and
10 slight but statistically significant increased liver weights at 2000 ppm. No treatment-related
11 histological effects were observed in the livers of animals exposed to less than 2000 ppm.

12 Several neurobehavioral tests were also conducted, and nine abnormalities were observed, one at
13 600 and 2000 ppm and eight at 2000 ppm, that could be associated with EDC exposure. These
14 effects included decreased resistance to removal from the home cage, increased palpebral
15 closure, increased lacrimation, decreased exterior thrust response, decreased response to sharp
16 noise and tail pinch, increased urination and defecation, and slight incoordination of gait.
17 Females exposed to 200 ppm had lower activity counts, but it was not statistically significant.
18 The lowest observed adverse effect level (LOEL) for neurological effects was 600 ppm.

19 The NOEL from this study was 50 ppm with a lowest observed effect level (LOEL) of 100 ppm
20 for degeneration and necrosis of the olfactory epithelium. Results indicate this NOEL is also
21 protective against kidney, liver, and neurological effects.

22 3.1.2.2.2 Supporting Animal Studies

23 Other animal studies evaluating the short-term effects of EDC inhalation are more limited (e.g.,
24 number of animals, endpoints examined, high doses used). However, the studies discussed below
25 are informative as supporting studies in the derivation of the 1-h acute ReV and ESL.

- 1 • Storer et al. (1984) compared the hepatotoxicity and genotoxicity following EDC exposure of
2 male C57BL/6 x C3HF₁ mice by inhalation, oral gavage, and intraperitoneal (i.p.) injection.
3 For the inhalation route, exposures were performed in a 30L stainless steel and glass
4 chamber, and EDC was vaporized in a gas washing bottle by a metered air flow.
5 Concentrations were monitored at 15 min intervals, with target concentrations of 150, 500,
6 1000, and 2000 ppm (time-weighted average concentrations of 158, 499, 1072, and 1946
7 ppm). For the gavage and i.p. routes, animals were treated with 200, 300, 400, 500, and 600
8 mg/kg EDC. All animals were exposed for 4 h and sacrificed 24 h later. The researchers
9 collected the livers and kidneys of all the animal and analyzed organ weights, and collected
10 blood to measure serum enzyme activities and blood urea nitrogen levels. Storer et al.
11 concluded that EDC is hepatotoxic and nephrotoxic by all three routes of exposure, with
12 threshold exposure levels at 500 ppm for inhalation, 400 mg/kg for i.p., and 500 mg/kg for
13 gavage. In the inhalation study, kidney-to-body weight ratios, serum L-iditol dehydrogenase,
14 and blood urea nitrogen levels were increased at 500 ppm. No differences were observed in
15 the 150 ppm exposure group compared to controls. The authors note that the route of
16 exposure plays an important role in the toxicity and genotoxicity of EDC.
- 17 • Zhang et al. (2011) exposed equal numbers of male and female Sprague-Dawley (SD) rats to
18 EDC concentrations of 0, 2500, 5000, and 10,000 mg/m³ (617, 1235, and 2470 ppm) for 6 h
19 and to 5000 mg/m³ for 0, 3, 6 and 12 h (6 rats/exposure group). Details on the exposure
20 methods were not provided. The researchers found that exposure to 5000 mg/m³ for a
21 minimum of 2 h caused an increase in water content in the cortex tissue in the rat brains.
22 Abnormal brain histopathology, including loose tissues and enlarged spaces around the cells,
23 was also observed after exposure to 5000 mg/m³. No significant differences were observed at
24 the lowest concentration tested; 2500 mg/m³ (617 ppm).
- 25 • Hepel et al. (1946) exposed mice, rats, guinea pigs, cats, dogs, and rhesus monkeys to target
26 EDC concentrations of 100, 200, 400, and 1000 ppm for 7 h/d, 5 d/wk, for varying lengths of
27 time. Details on the strains/breeds were not provided, and not all species were exposed to
28 every concentration. All of the animals were exposed in a 4'x4'x6' chamber and EDC was
29 volatilized by passing a steady flow of air through the liquid. EDC concentrations were
30 calculated using the air flow rate and the rate of volatilization, and measurements were taken
31 daily through gas samples from the chamber. The analytical concentrations were 0.42, 0.73,
32 1.54, 3.9 mg/L (420, 730, 1540, and 3900 mg/m³, respectively). At 1000 ppm, high mortality
33 rates were seen in the rats, rabbits, and guinea pigs after only a few exposures. The cats and
34 dogs were more resistant, but eventually died at 1000 ppm EDC. One monkey died after the
35 second exposure, while a second monkey survived for 43 exposures. Autopsy of the animals
36 revealed lung congestion, fatty degeneration in the liver, and fatty changes in the liver.
37 Mortality was also seen in the 200 and 400 ppm groups; however more animals survived a
38 greater number of exposures than in the 1000 ppm group. Similar causes of death were
39 determined after autopsy. At 100 ppm, all 16 female and 23 male rats survived at least 74
40 exposures. Breeding of the 16 female rats during the exposure period resulted in pregnancies
41 in all but one of the rats. Survival records of the pups did not appear to be affected. Two
42 guinea pigs died after 20 and 69 exposures, but an unusually high mortality rate in the

1 controls animals made this data unreliable. All 19 mice exposed repeatedly to 100 ppm also
2 survived, and no gross effects were observed.

- 3 • Wang et al. (2013) exposed female albino mice via inhalation to nominal EDC
4 concentrations of 225, 450, and 900 mg/m³ (56, 111, and 222 ppm respectively) for 3.5 h/d
5 for 10 d. EDC was placed on filter paper on a plate suspended in a 100 L chamber, and eight
6 mice were exposed concurrently. Two hours after the final exposure, open field tests were
7 performed, after which the animals were sacrificed and their brains examined. Several
8 behavioral changes were observed, including a significant and dose-dependent decrease in
9 the number of line crossings at 450 and 900 mg/m³, a significant increase in vertical activity
10 at 225 mg/m³ that was not dose-dependent, an increase in time spent in the central zone that
11 was not significant or dose-dependent, and a significant increase in the production of urine.
12 Examination of the brains revealed higher levels of malondialdehyde in animals exposed to
13 225 mg/m³ EDC, and higher levels of superoxide dismutase in animals exposed to 0.9 g/m³
14 EDC, but neither response showed a dose-dependent trend. The researchers also found
15 significantly higher levels of nitric oxide at 450 mg/m³ EDC and inducible nitric oxide
16 synthase at 225 mg/m³ EDC, suggestive of damage from reactive nitrogen species. The
17 lowest observed effect level (LOEL) from this study was 225 mg/m³ EDC (56 ppm) for
18 enzyme changes in the brain and 450 mg/m³ (111 ppm) for neurobehavioral changes.
- 19 • Igwe et al. (1986) exposed male Sprague-Dawley (SD) rats to 150, 300, and 450 ppm EDC
20 continuously for 30 d (24 h/d, 12 rats/exposure group). Animals were treated in stainless steel
21 and glass exposure chambers, and EDC vapors were generated by pumping liquid EDC
22 through an air feed. Concentrations were monitored continuously and recorded hourly, with
23 mean average chamber concentrations of 153, 304, and 455 ppm. Animals were sacrificed on
24 day 31, and body and liver weights were recorded. Blood samples were taken to analyze for
25 enzymes that are representative of liver function. Increased liver-to-body weight ratios and
26 increased 5-nucleotidase activity were observed following exposure to 450 ppm EDC. At 300
27 ppm, exposed rats had statistically significant increases in hepatic protein content and
28 glutathione levels and decreased hepatic cytochrome P₄₅₀ content. At 150 ppm, exposed rats
29 also had statistically significant increases in hepatic protein content; however this trend was
30 not dose-dependent.

31 ***3.1.2.3 Reproductive and Developmental Studies***

32 Studies regarding the reproductive and developmental effects of EDC are limited. A single
33 inhalation study on the possible reproductive effects of EDC in humans is available and is
34 detailed in ATSDR (2001).

- 35 • Zhao et al. (1989) observed an increase in the rates of premature birth in female workers and
36 the wives of male workers at a Chinese synthetic factory that were exposed to EDC.
37 Concentrations ranged from 0.4 to 384 ppm at two locations, and around 100 workers were
38 included in the study. Workers were exposed either throughout their pregnancy (female
39 workers) or for at least one year prior to their wife giving birth (male workers). No

1 information was given on the other chemical exposures that may have also occurred, or any
2 other physical/environmental confounders that may have also been present.

3 Several animal inhalation studies have examined the reproductive and developmental effects of
4 EDC, although the exposure durations span multiple days. ATSDR (2001) concluded that “the
5 overall evidence from inhalation studies in rats and rabbits indicate that EDC is not a
6 developmental toxicant”.

- 7 • Rao et al. (1980), as described in USEPA (2010), exposed pregnant SD rats and White
8 New Zealand rabbits to 0, 100, or 300 ppm EDC for 7 h/d on gestational days (GD) 6-15
9 (rats) or 6-18 (rabbits). Details on the exposure method were not given. Animals were
10 sacrificed on GD 21 (rats) or 29 (rabbits), and the uteri and fetuses were examined.
11 Maternal mortality occurred at 100 ppm in rabbits and 300 ppm in rats. None of the
12 fetuses from rats exposed to 300 ppm survived, possibly due to maternal toxicity.
13 Exposure to 100 ppm in rats did not affect the mean litter size, the incidence of
14 resorptions, fetal body measurements, or sex ratio. Exposure to 100 and 300 ppm in
15 rabbits did not affect the rate of pregnancy, number of implantation sites, resorption
16 incidence, litter size, sex ratio, or fetal measurements in surviving rabbits.
- 17 • Rao et al. (1980), as described in USEPA (2010), exposed groups of SD rats to 0, 25, 75,
18 and 100 ppm (20/sex/group with controls at 30/sex) for 6 h/d, 5 d/wk for 60 d. Rats were
19 mated after the initial 60 d exposure, and exposure was continued through gestation,
20 discontinued from GD 21 to postnatal day (PND) 4, and then continued until the second
21 breeding cycle. The first sets of pups were sacrificed at PND 21 and the adult animals
22 were allowed to mate again. All animals were sacrificed once the second set of pups
23 reached PND 21. No treatment-related abnormalities in fertility, reproduction, or fetal
24 development were observed. There were no exposure-related changes in fetal growth,
25 organ weight or histology.
- 26 • Zhao et al. (1989), as described in USEPA (2010) and ATSDR (2001), exposed pregnant
27 Wistar rats to 0, 6.1, or 51.3 ppm EDC for 6 h/d from 2 wk prior to mating until GD 20.
28 The exact duration of exposure is unknown. No effects were observed in the maternal
29 endpoints examined, including body weight gain, impregnation rates, blood cell counts,
30 blood protein content, and urine protein. However preimplantation loss was significantly
31 increased compared to controls (31.0% versus 10.2%) in the highest exposure group
32 (51.3 ppm). No fetal effects were observed at either dose. The USEPA (2010) and
33 ATSDR (2001) considered these results questionable because of difficulties translating
34 the information from Chinese, including inconsistencies regarding species information
35 and the number of animals used, and because statistical analyses were not conducted.

36 A recent one-generation drinking water study was conducted examining possible reproductive
37 effects of EDC in mice and rats (Charlap 2015). This study was conducted as part of the
38 Hazardous Air Pollutants (HAP) Task Force and submitted to the USEPA. A second study

1 reported on a physiologically based pharmacokinetic (PBPK) model used to extrapolate the oral
2 doses to inhalation exposures (Sweeney and Gargas, 2015).

3 • Charlap (2015) exposed groups of F₀ and F₁ male and female CrI:CD(SD) rats
4 (27/sex/group) to EDC in drinking water at target concentrations of 0, 50, 150, and
5 300 mg/kg/d. Actual EDC doses were less than targeted doses in most groups due to
6 what the authors suspected was palatability of the water. F₀ males were exposed
7 throughout mating until euthanasia (day 92 or 93), while F₀ females continued to
8 receive EDC doses throughout mating, gestation, and lactation (lactation day 22).
9 Following mating of F₀ males and females, F₁ offspring were selected on PND 21 in
10 groups of 20/sex/group and continued to receive EDC until euthanasia. An expansive
11 set of parameters were recorded for all the animals, including clinical observations,
12 body weights, food and water consumption, clinical pathology tests, complete
13 necropsies, and reproductive analyses. Lower mean body weights, body weight gains,
14 and food and water consumption were noted in both the F₀ and F₁ groups. These
15 decreases were attributed to the decreased palatability of EDC in the drinking water.
16 No signs of reproductive toxicity were observed in any of the groups tested, resulted
17 in a reproductive NOAEL of 300 mg/kg/d (actual exposure level of 155 mg/kg/d for
18 F₀ males, 182 mg/kg/d for F₀ females, 184 mg/kg/d for F₁ males, and 169 mg/kg/d for
19 F₁ females). Sweeney and Gargas (2015) determined that the lowest NOAEL of 155
20 mg/kg/d was equivalent to a continuous inhalation exposure of 62 ppm.

21 All of the available reproductive and developmental data are based on multiple exposures, with
22 the lowest reported effects observed at 51.3 ppm in the Zhao et al. (1989) study, which was
23 deemed by the USEPA (2010) as unreliable. Using a Point of departure (POD) of 50 ppm from a
24 single 8 h exposure should protect against any possible reproductive or developmental effects
25 that may occur at higher or more frequent doses. It should be noted that acute EDC exposure has
26 a very steep dose-response curve with respect to mortality (OEHHA 2000), as seen in the animal
27 studies described above.

28 **3.1.3 Mode-of-Action (MOA) Analysis and Dose Metric**

29 Absorption, distribution, and metabolism of EDC following inhalation or ingestion are rapid and
30 complete in rats, with 85% of the metabolites being excreted in urine (OEHHA 2000). Acute
31 inhalation of EDC at relatively high concentrations has been shown to cause CNS, liver, and
32 kidney effects. At low concentrations, EDC has been shown to cause POE effects including
33 degeneration and necrosis of nasal olfactory epithelium. The mechanism by which EDC acts at
34 the POE or systemically is not well understood. Animal studies suggest that EDC may be
35 metabolized to 2-chloroacetaldehyde and other reactive metabolites, which are able to bind and
36 inhibit cellular macromolecules (ASTDR 2001). Exposure to the parent compound is the only
37 available dose metric.

1 **3.1.4 Point of Departure (POD) for Key Animal Study and Critical Effects**

2 Hotchkiss et al. (2010) observed very slight degeneration and necrosis of the olfactory
3 epithelium in 4 out of 10 rats exposed to 100 ppm EDC for 8 h. The NOEL from this study was
4 50 ppm, which will be conservatively used at the POD in further calculations of the 1-h acute
5 ReV and ^{acute}ESL.

6 **3.1.4.1 Default Exposure Duration Adjustments**

7 The 8-h duration (C_1) in the key study by Hotchkiss et al. (2010) was adjusted to a POD_{ADJ} of 1-
8 h exposure duration (C_2) using Haber's Rule as modified by ten Berge et al. (1986) ($C_1^n \times T_1 =$
9 $C_2^n \times T_2$) with $n = 3$, where both concentration and duration play a role in toxicity:

$$\begin{aligned} 10 \quad C_2 &= [(C_1)^3 \times (T_1 / T_2)]^{1/3} \\ 11 \quad &= [(50 \text{ ppm})^3 \times (8 \text{ h} / 1 \text{ h})]^{1/3} \\ 12 \quad &= 100 \text{ ppm} = POD_{ADJ} \end{aligned}$$

13 **3.1.4.2 Default Dosimetry Adjustments from Animal-to-Human Exposure**

14 EDC is very water soluble and causes POE effects (Category 1 gas) at low concentrations and
15 acts systemically (i.e., as a Category 3 gas) on the CNS, liver, and kidneys at high
16 concentrations. The critical effect of very slight degeneration and necrosis of the olfactory
17 epithelium would suggest using a pharmacokinetic dosimetric animal-to-human adjustment
18 factor (DAF) of 1 as a Category 1 gas acting on the extrathoracic region (ET). Likewise, in
19 regard to the systemic effects occurring at higher concentrations, the default pharmacokinetic
20 animal-to-human dosimetric adjustment for a Category 3 gas is a blood:gas partition coefficient
21 animal/human ratio of 1 (TCEQ 2012). Thus, all these considerations support using a dosimetric
22 animal-to-human adjustment of 1 for the POD_{ADJ} (i.e., use a DAF_{ET} of 1). Thus, the POD_{HEC} is
23 equal to the POD_{ADJ} of 100 ppm.

24 **3.1.5 Adjustments to the POD_{HEC}**

25 The POD_{HEC} based on a NOEL from the Hotchkiss et al. (2010) study was used as the POD and
26 UFs were applied to derive the 1-h acute ReV (i.e., assume a threshold MOA). The following
27 uncertainty factors (UFs) were applied to the POD_{HEC} of 100 ppm: 10 for intraspecies variability
28 (UF_H), 3 for extrapolation from animals to humans (UF_A), and 6 for database uncertainty (UF_D),
29 for a total UF of 180.

- 30 • An UF_H of 10 was used to account for variation in sensitivity among the members of the
31 human population including possible child/adult differences, those with pre-existing medical
32 conditions, etc.;
- 33 • An UF_A of 3 was used to account for potential pharmacodynamic differences between
34 animals and humans (pharmacokinetic adjustment was already performed); and

- 1 • An UF_D of 6 was used because although there are several acute studies in multiple species
2 available for EDC, including reproductive and developmental studies, only a single study
3 looked at possible nasal/respiratory effects, and additional studies could provide more insight
4 into this target region. Also, EDC shows a very steep dose response curve (mild respiratory
5 effects from a single exposure to 100 ppm and death after multiple exposures to 100 ppm),
6 which requires due consideration when selecting the UF_D given the studies available. The
7 quality of the study used as the POD is considered high, and the confidence in the acute
8 database is medium to high.

9 acute ReV = $POD_{HEC} / (UF_H \times UF_A \times UF_D)$
10 = 100 ppm / (10 x 3 x 6)
11 = 100 ppm / 180
12 = 0.5555 ppm
13 = 555.5 ppb or 550 ppb (rounded to two significant digits)

14 **3.1.6 Health-Based 1-h Acute ReV and ^{acute}ESL**

15 In deriving the acute 1-h ReV for EDC, no numbers were rounded between equations until the
16 ReV was calculated. Once the ReV was calculated, it was rounded to two significant figures. The
17 resulting 1-h acute ReV is 550 ppb (2200 $\mu\text{g}/\text{m}^3$) based on the Hotchkiss et al. (2010) study. The
18 rounded acute ReV was then used to calculate the ^{acute}ESL. At the target hazard quotient (HQ) of
19 0.3, the ^{acute}ESL is 160 ppb (650 $\mu\text{g}/\text{m}^3$) (Table 6).

20

1 **Table 6. Derivation of the 1-h Acute ReV and ^{acute}ESL**

Parameter	Values and Descriptions
Study	Hotchkiss et al. (2010)
Study Population	Male and female Fischer 344 rats, 5/sex/group in two acute inhalation toxicity studies and 10/sex/group in an acute neurotoxicity study
Study Quality	High
Exposure Method	Exposure via inhalation at 0, 50, 100, 150, 200, 600 and 1000 ppm for 4 or 8 h
Critical Effect	Sight degeneration and necrosis of olfactory epithelium
POD (NOEL)	50 ppm
Exposure Duration	8 h
Extrapolation from 8 h to 1 h (POD _{ADJ})	100 ppm
POD _{HEC} (1 h)	100 ppm
Total UF	180
<i>Intraspecies UF</i>	10
<i>Interspecies UF</i>	3
<i>Incomplete Database UF</i> <i>Database Quality</i>	6 Medium to high
acute ReV [1 h] (HQ = 1)	550 ppb (2200 µg/m ³)
^{acute}ESL [1 h] (HQ = 0.3)	160 ppb (650 µg/m ³)

2 **3.2 Health-Based 24-h ReV**

3 **3.2.1 Background on 24-Hour AMCVs**

4 For chemicals detected in the ambient air monitoring network, short-term AMCVs have
 5 generally been derived by the TCEQ to evaluate 1-h reported concentrations and long-term
 6 AMCVs have been derived to evaluate annual averages. Since a significant amount of ambient
 7 air data is collected over a 24-h duration, the derivation of chemical-specific 24-h AMCV values
 8 is needed to better evaluate ambient 24-h data. This consideration applies to EDC since it is
 9 detected in the TCEQ ambient air monitoring network and toxicity data are available to derive a
 10 24-h ReV. Without a 24-h AMCV for EDC, only a limited evaluation of the reported 24-h levels

1 is possible because 1-h and chronic (i.e., lifetime) AMCVs are generally inappropriate for this
2 purpose. Thus, the development of a 24-h AMCV is necessary for the best possible health effects
3 evaluation of individual 24-h sample results, and would significantly complement the short-term
4 and chronic evaluations of EDC in ambient air data.

5 **3.2.2 Key and Supporting Studies**

6 The key and supporting studies listed in Section 3.1 are the same studies used in the derivation of
7 the 24 h ReV. Hotchkiss et al. (2010) will be used as the key study, as discussed in Section
8 3.1.2.2 above.

9 **3.2.3 MOA and Dose Metric**

10 Absorption, distribution, and metabolism of EDC following inhalation or ingestion are rapid and
11 complete in rats, with 85% of the metabolites being excreted in urine (OEHHA 2000). Acute
12 inhalation of EDC at relatively high concentrations has been shown to cause CNS, liver, and
13 kidney effects. At low concentrations, EDC has been shown to cause POE effects including
14 degeneration and necrosis of nasal olfactory epithelium. The mechanism by which EDC acts at
15 the POE or systemically is not well understood. Animal studies suggest that EDC may be
16 metabolized to 2-chloroacetaldehyde and other reactive metabolites, which are able to bind and
17 inhibit cellular macromolecules (ASTDR 2001). Exposure to the parent compound is the only
18 available dose metric.

19 **3.2.4 PODs for Key Study, Critical Effects and Dosimetric Adjustments**

20 Hotchkiss et al. (2010) observed very slight degeneration and necrosis of the olfactory
21 epithelium in 4 out of 10 rats exposed to 100 ppm EDC for 8 h. The NOEL from this study was
22 50 ppm, which will be conservatively used at the POD in further calculations of the acute 24-h
23 ReV.

24 ***3.2.4.1 Default Exposure Duration Adjustments***

25 The 8-h duration (C_1) in the key study by Hotchkiss et al. (2010) was adjusted to a POD_{ADJ} of
26 24-h exposure duration (C_2) using Haber's Rule as modified by ten Berge et al. (1986) ($C_1^n \times T_1$
27 = $C_2^n \times T_2$) with $n = 1$, where both concentration and duration play a role in toxicity:

$$\begin{aligned} 28 \quad C_2 &= [(C_1) \times (T_1 / T_2)] \\ 29 \quad &= [(50 \text{ ppm}) \times (8 \text{ h} / 24 \text{ h})] \\ 30 \quad &= 16.6667 \text{ ppm} = POD_{ADJ} \end{aligned}$$

31 ***3.2.4.2 Default Dosimetry Adjustments from Animal-to-Human Exposure***

32 EDC is very water soluble and acts as a POE irritant (Category 1 gas) at low concentrations and
33 acts systemically (i.e., as a Category 3 gas) on the CNS, liver, and kidneys at high
34 concentrations. The critical effect of very slight degeneration and necrosis of the olfactory

1 epithelium would suggest using a pharmacokinetic dosimetric animal-to-human adjustment
2 factor (DAF) of 1 as a Category 1 gas acting on the ET region. Likewise, in regard to the
3 systemic effects occurring at higher concentrations, the default pharmacokinetic animal-to-
4 human dosimetric adjustment for a Category 3 gas is a blood:gas partition coefficient
5 animal/human ratio of 1 (TCEQ 2012). Thus, all these considerations support using a dosimetric
6 animal-to-human adjustment of 1 for the POD_{ADJ} (i.e., use a DAF_r of 1). Thus, the POD_{HEC} is
7 equal to the POD_{ADJ} of 16.6667 ppm.

8 **3.2.5 Adjustments to the POD_{HEC}**

9 The POD_{HEC} based on a NOEL from the Hotchkiss et al. (2010) study was used as the POD and
10 UFs were applied to derive the 24-h ReV (i.e., assume a threshold MOA for a noncarcinogenic
11 endpoint). The following uncertainty factors (UFs) were applied to the POD_{HEC} of 16.6667 ppm:
12 10 for UF_H , 3 for UF_A , and 6 for UF_D , for a total UF of 180.

- 13 • An UF_H of 10 was used to account for variation in sensitivity among the members of the
14 human population including possible child/adult differences, those with pre-existing medical
15 conditions, etc.;
- 16 • An animal-to-human UF_A of 3 was used to account for potential pharmacodynamic
17 differences between animals and humans (pharmacokinetic adjustment was already
18 performed); and
- 19 • A database deficiency UF_D of 6 was used because although there are several acute studies in
20 multiple species available for EDC, including reproductive and developmental studies, only a
21 single study looked at possible nasal/respiratory effects, and additional studies could provide
22 more insight into this target region. Also, EDC shows a very steep dose response curve (mild
23 respiratory effects at 100 ppm and death in some animals at 300 ppm), which requisites due
24 consideration when selecting the UF_D given the studies available. The quality of the study
25 used as the POD is considered high, and the confidence in the acute database is medium to
26 high.

$$\begin{aligned} 27 \quad \text{acute ReV} &= POD_{HEC} / (UF_H \times UF_A \times UF_D) \\ 28 &= 16.6667 \text{ ppm} / (10 \times 3 \times 6) \\ 29 &= 16.6667 \text{ ppm} / 180 \\ 30 &= 0.0926 \text{ ppm} \\ 31 &= 92.6 \text{ ppb or } 93 \text{ ppb (rounded to two significant digits)} \end{aligned}$$

32 **3.2.6 Health-Based 24-h ReV**

33 In deriving the acute 24-h ReV for EDC, no numbers were rounded between equations until the
34 ReV was calculated. Once the ReV was calculated, it was rounded to two significant figures. The
35 resulting 24-h acute ReV is 93 ppb ($380 \mu\text{g}/\text{m}^3$) based on the Hotchkiss et al. (2010) study (Table
36 7).

1 **Table 7. Derivation of the 24-h Acute ReV**

Parameter	Values and Descriptions
Study	Hotchkiss et al. (2010)
Study Population	Male and female Fischer 344 rats, 5/sex/group in two acute inhalation toxicity studies and 10/sex/group in an acute neurotoxicity study
Study Quality	High
Exposure Method	Exposure via inhalation at 0, 50, 100, 150, 200, 600 and 1000 ppm for 4 or 8 h
Critical Effect	Degeneration and necrosis of olfactory epithelium
POD (NOEL)	50 ppm
Exposure Duration	8 h
Extrapolation from 8 h to 24 h (POD _{ADJ})	16.6667 ppm
POD _{HEC} (24 h)	16.6667 ppm
Total UF	180
<i>Intraspecies UF</i>	10
<i>Interspecies UF</i>	3
<i>Incomplete Database UF</i>	6
<i>Database Quality</i>	Medium to high
acute ReV [24 h] (HQ = 1)	93 ppb (380 µg/m³)

2

3 **3.3 Welfare-Based Acute ESLs**

4 **3.3.1 Odor Perception**

5 EDC has a pleasant odor and a sweet taste (ATSDR 2001). Published odor detection threshold
6 values are summarized in Table 8 (TCEQ 2015).

7

1 **Table 8. Accepted Odor Studies Conducted for EDC**

Investigator	Odor Detection Threshold Value
Hellman (1974)	24,000 $\mu\text{g}/\text{m}^3$ (6,000 ppb)
May (1966)	450,000 $\mu\text{g}/\text{m}^3$ (110,000 ppb)

2 Because these odor values are significantly higher than the determined acute ESLs, and the odor
3 of EDC is described as pleasant and sweet, an ^{acute}ESL_{odor} will not be derived (TCEQ 2015).

4 **3.3.2 Vegetation Effects**

5 After a literature review, there was no data found on any adverse effects of EDC on vegetation.

6 **3.4 Short-Term ESL and Values for Air Monitoring Evaluation**

7 The acute evaluation resulted in the derivation of the following values:

- 8 • Acute 1-h ReV = 550 ppb (2,200 $\mu\text{g}/\text{m}^3$)
- 9 • ^{acute}ESL [1 h] = 160 ppb (650 $\mu\text{g}/\text{m}^3$)
- 10 • Acute 24-h ReV = 93 ppb (380 $\mu\text{g}/\text{m}^3$)

11 For the evaluation of ambient air monitoring data, the acute 1-h ReV for EDC is 550 ppb (2,200
12 $\mu\text{g}/\text{m}^3$) (Table 6), and the acute 24-h ReV is 93 ppb (380 $\mu\text{g}/\text{m}^3$) (Table 7). The short-term ESL
13 used for air permit reviews is the health-based ^{acute}ESL of 160 ppb (650 $\mu\text{g}/\text{m}^3$) (Table 2).

14 **3.5 Acute Inhalation Observed Adverse Effect Level**

15 Risk assessors, and the general public, often ask to have information on the levels in air where
16 health effects would be expected to occur. So, when possible, the TCEQ provides chemical-
17 specific observed adverse effects levels in DSDs (TCEQ 2012). As the basis for development of
18 inhalation observed adverse effect levels is limited to available data, future studies could
19 possibly identify a lower POD for this purpose. Regarding critical effects due to acute EDC
20 exposure, the animal study by Hotchkiss et al. (2010) found a 8-h rat LOEL of 100 ppm for nasal
21 effects. This animal LOEL was used as the animal acute inhalation observed adverse effect level
22 for extrapolation to humans. No duration adjustment was made (TCEQ 2012). As discussed in
23 Section 3.1.5.2, for these effects the animal-to-human dosimetric adjustment results in a
24 LOEL_{HEC} equal to the animal exposure concentration (e.g., a DAF of 1 is used). Thus, the 8-h
25 LOEL_{HEC} based on this animal study is estimated to be 100 ppm. This value is applicable to both
26 the 1-h and 24-h ReV.

27 The LOEL_{HEC} determined from an animal study represents a concentration at which it is possible
28 that similar effects could occur in some individuals exposed to this level over the same duration
29 as used in the study (8 h) or longer. Importantly, effects are not a certainty due to potential

1 interspecies and intraspecies differences in sensitivity. The acute inhalation observed adverse
2 effect level of 100 ppm (400 $\mu\text{g}/\text{m}^3$) is provided for informational purposes only (TCEQ 2012).

3 The margin of exposure between the estimated acute inhalation observed adverse effect level of
4 100 ppm (400 $\mu\text{g}/\text{m}^3$) and the acute 1-h ReV of 550 ppb is a factor of 180 and the 24-h ReV of
5 93 ppb is a factor of over 1000.

6 **Chapter 4 Chronic Evaluation**

7 ***4.1 Noncarcinogenic Potential***

8 Data on the chronic toxicity of EDC, other than those from carcinogenicity studies, are limited.
9 Unlike acute exposures, long-term studies tend to show negative results at concentrations that do
10 not also induce mortality. The toxicological profiles of EDC from the USEPA (2010), ATSDR
11 (2001), and IARC (1999) were reviewed for this section along with conducting a literature
12 review for any more current studies. Because of the insufficient nature of the human data, an
13 animal study with the appropriate UFs will be used to derive the chronic ReV.

14 **4.1.2 Physical/Chemical Properties**

15 The primary physical and chemical properties of EDC are discussed in Chapter 3 and
16 summarized in Table 3.

17 **4.1.3 Key and Supporting Studies**

18 ***4.1.3.1 Human Studies***

19 Several occupational studies have looked at workers exposed to EDC, but unfortunately these
20 data are not very informative due to either co-exposures to other hazardous chemicals, such as
21 vinyl chloride, insufficient exposure data, and/or lack of a dose-response relationship. A few of
22 these studies can be found in the USEPA (2010) and ATSDR (2001) toxicological reviews of
23 EDC and are detailed in in Section 3.1.2.1.

24 Due to the lack of sufficient human data, animal data were used to develop the chronic ReV.

25 ***4.1.3.2 Animal Studies***

26 **4.1.3.2.1 Key study**

27 Spreafico et al. (1980) and Maltoni et al. (1980), as detailed in USEPA (2010) and OEHHA
28 (2000), exposed male and female SD rats and Swiss mice (90/sex/group) to inhalation of 5, 10,
29 50, or 150-250 ppm EDC for 7 h/d, 5 d/wk, for up to 18 months. Spreafico et al. (1980) reported
30 on the methods and chronic toxicity of EDC, while Maltoni et al. (1980) reported the neoplastic
31 endpoints. Control groups consisted of 180 rats (chamber control and untreated) and 249 mice
32 (untreated only). Originally the highest exposure concentration was set at 250 ppm, but due to

1 high toxicity in the animals after several days, the concentration was reduced to 150 ppm. Body
2 weights were recorded every 2 wks, and 8-10 animals from each group were sacrificed at 3, 6,
3 and 18 months. A series of parameters were examined, including hematology, serum chemistry,
4 urinalysis, gross necropsy, and microscopic examination of the major tissues, including liver,
5 lungs, kidneys, and gonads.

6 No non-neoplastic histology findings were reported, and no information was given as to the
7 toxicity observed at 250 ppm. No exposure-related mortality was reported at 150 ppm. Several
8 changes in hematology, serum chemistry, and urinalysis were statistically significant, but none
9 showed a dose- or temporal-response. A second group of 8-10, 14-month old rats was exposed to
10 50 ppm for 12 months and they showed significant and dose-dependent increases in in ALT and
11 uric acid in the serum. This effect was not observed in the rats exposed to the same concentration
12 for 18 months, which the authors suggest could be because they were younger when treatment
13 began (3 months old). No differences in any of the parameters examined were observed
14 following 18-month exposure to 5 or 10 ppm. USEPA determined that the LOAEL from this
15 study was 50 ppm for suggestive liver and kidney toxicity (increased ALT and uric acid,
16 respectively) with a NOAEL of 10 ppm.

17 **4.1.3.2.2 Supporting studies**

18 Several studies have looked at chronic exposure of laboratory animals to EDC, although there
19 have not been many significant findings at concentrations that do not also induce mortality.
20 Several of these studies are summarized in USEPA (2010) and OEHHA (2000), and brief
21 descriptions are provided below.

- 22 • Nagano et al. (2006) exposed male and female specific pathogen free (SPF) F344/DuCrj rats
23 and Crj:BDF1 mice to EDC via inhalation at concentrations of 0, 10, 40, and 160 ppm (rats)
24 and 0, 10, 30, and 90 ppm (mice) for 2 years (yr) (50/sex/exposure group). Concentrations
25 were determined from an initial 13-wk study where rats and mice showed high mortality
26 rates when exposed to 320 and 160 ppm, respectively, but no overt toxicity at 160 and 80
27 ppm. EDC was vaporized by bubbling clean air through the liquid in a temperature-regulated
28 glass flask, and exposures were conducted in a 7600 L chamber for rats and a 3700 L
29 chamber for mice. Chamber concentrations were measured every 15 min, with 2-yr averages
30 of 10, 39.8, and 159.7 ppm for the rats, and 10, 30, and 89.8 ppm for the mice. Body weights
31 and food consumption were monitored once a week for the first 14 wk, then once a month for
32 the remaining two years. A number of non-neoplastic endpoints were examined, including
33 survival, urinary parameters, hematology and blood chemistry, and organ weight and gross
34 pathology. Microscopic evaluations of neoplastic lesions were also conducted and are
35 discussed in Section 4.2.2.2. No differences were observed in the survival rates, growth rates,
36 or food consumption in any of the exposed rats compared to control. Female mice exposed to
37 30 ppm EDC had significantly lower survival rates than the control animals, but there was no
38 difference in the survival rate of the exposed male mice. Since this decrease in survival was
39 not dose-dependent, the authors concluded that it was not exposure-related. No differences

1 were observed in the growth rates or food consumption in any of the exposed mice compared
2 to control. No exposure-related differences were observed in any hematological, blood
3 biochemical or urinary parameter in any of the exposed animals. No exposure-related, non-
4 neoplastic lesions were observed in any of the EDC-treated animals. The free-standing
5 NOAEL from this study for non-neoplastic changes is 160 ppm for rats and 90 ppm for mice.

- 6 • Cheever et al. (1990) exposed groups of 50 SD [CrI:CD(SD)BR outbred] rats to 50 ppm EDC
7 for 7 h/d, 5 d/wk for 2 yr (except for holidays). Animals were exposed in 2.2 m³ stainless
8 steel and glass chambers. EDC was volatilized by passing compressed air through liquid
9 EDC. Nominal concentrations were calculated on a daily basis, and analytical concentrations
10 were measured hourly. The average chamber concentration was 50.4 ppm for the target
11 concentration of 50 ppm. Body weights, food and water consumption were measured weekly.
12 Animals that either died during the study or were sacrificed after 2 yr were examined for
13 gross histopathological changes and blood parameters related to the metabolism of EDC.
14 Exposure to 50 ppm had no effect on mortality, body weight, food or water consumption, or
15 the appearance of the animals. Female rats had a slight increase in basophilic focal cellular
16 changes in the pancreas, but this was not observed in the male rats.
- 17 • Hofmann et al. (1971, as cited in USEPA 2010) exposed cats (2/sex/group), rabbits
18 (2/sex/group), guinea pigs (5/sex/group), and SD rats (5/sex/group) to 0, 100, or 500 ppm
19 EDC for 6 h/d, 5 d/wk for up to 17 wk. The researchers looked at a number of endpoints
20 including body and organ weights, hematology, urinalysis, serum chemistry, liver function,
21 and microscopic examination of specific organs. However not all of the parameters were
22 evaluated for each species. At 500 ppm, 3/4 rabbits died after 10-17 exposures, 9/10 guinea
23 pigs died after 4-14 exposures, and all the rats died after 1-5 exposures. At 100 ppm, there
24 were no exposure-related differences in clinical signs, clinical chemistry, body weights, liver
25 or kidney weights, or histopathology. The USEPA noted a NOAEL for this study of 100
26 ppm.
- 27 • Spencer et al. (1951) exposed male and female Wistar rats, guinea pigs, albino rabbits, and
28 rhesus monkeys to 0, 100, 200, and 400 ppm EDC for 7 h/d, 5 d/wk for up to 35 weeks. The
29 animals were exposed in a metal chamber, and EDC vapor was generated by passing air
30 through a vaporizer containing liquid EDC. Chamber concentrations were measured
31 continuously, and although the analytical concentrations were not provided, the authors state
32 that “in every case the vapor was uniformly held within 10% of the desired concentration.”
33 Animals were examined for several toxicity endpoints including body and organ weights,
34 gross organ histology, hematology, and serum chemistry. At 400 ppm, significant mortality
35 occurred in all of the species tested, with the rats surviving no more than 40 exposures,
36 guinea pigs no more than 24 exposures, and the monkeys no more than 12 exposures. Three
37 rabbits survived 165 exposures with no changes in the toxicity parameters examined. At 200
38 ppm, all of the exposed rats and guinea pigs survived 151 and 180 exposures, respectively;
39 they were only the species tested at this concentration. The exposed rats showed no signs of
40 toxicity on body and organ weights, gross organ histology, hematology, and serum
41 chemistry. Male and female guinea pigs had smaller body weights throughout the study, but
42 only the male guinea pigs were significantly smaller than their control counterparts at the end

1 of the study. The authors noted that about half of the guinea pigs showed slight
2 parenchymatous degeneration of the liver, dispersed fat vacuoles, and a slight increase in
3 total lipid, phospholipid, fat, and cholesterol compared to controls. No other effects were
4 observed. At 100 ppm, all of the exposed rats and guinea pigs survived 151 and 121
5 exposures, respectively. No changes were observed in any of the toxicity parameters tested.
6 The NOAEL for this study is 100 ppm with a LOAEL for liver function changes and
7 decreased body weight at 200 ppm.

8 **4.1.3.3 Reproductive and Developmental Studies**

9 Studies regarding the potential reproductive and developmental effects of EDC in humans are
10 limited. A single inhalation study on the possible reproductive effects of EDC in humans is
11 available and is detailed in ATSDR (2001) and detailed in Section 3.1.2.3.

12 All of the available reproductive and developmental data are based on multiple exposures, with
13 the lowest reported effects observed at 51.3 ppm in the Zhao et al. (1989) study, which was
14 deemed unreliable by the USEPA (2010). Regardless, this result is similar to the LOAEL (50
15 ppm) from the key animal study (Spreafico et al. 1980), and using a POD of 10 ppm with
16 appropriate UFs to derive the chronic ReV should also protect against potential reproductive or
17 developmental effects that may occur at higher doses (e.g., >100 ppm in Rao et al. 1980).

18 **4.1.4 MOA and Dose Metric**

19 Absorption, distribution, and metabolism of EDC following inhalation or ingestion are rapid and
20 complete in rats, with 85% of the metabolites being excreted in urine (OEHHA 2000). Chronic
21 inhalation studies tend to show little to no effects at doses that do not also result in mortality.
22 Liver and kidney changes suggest that chronic EDC exposure acts systemically, although the
23 mechanism is not well understood. Animal studies suggest that EDC may be metabolized to 2-
24 chloroacetaldehyde and other reactive metabolites, which are able to bind and inhibit cellular
25 macromolecules (ASTDR 2001). However, the specific MOA for chronic EDC toxicity remains
26 unknown. Exposure to the parent compound is the only available dose metric.

27 **4.1.5 PODs for Key Study, Critical Effects and Dosimetric Adjustments**

28 Based on the key study presented above (Spreafico et al. 1980), the TCEQ identifies 10 ppm (40
29 mg/m³) as the NOAEL and POD based on rat liver and kidney toxicity.

30 **4.1.5.1 Default Exposure Duration Adjustments**

31 Animals were exposed for 7 h/day, 5 d/wk, for up to 18 months. An adjustment from a
32 discontinuous to a continuous exposure duration was conducted (TCEQ 2012) as follows:

$$33 \quad \text{POD}_{\text{ADJ}} = \text{POD}_{\text{HEC}} \times (\text{D}/24 \text{ h}) \times (\text{F}/7 \text{ d})$$

34 where:

35 D = Exposure duration, hours per day

1 F = Exposure frequency, days per week

2 $POD_{ADJ} = 10 \text{ ppm} \times (7/24) \times (5/7) = 2.0833 \text{ ppm}$

3 **4.1.5.2 Default Dosimetry Adjustments from Animal-to-Human Exposure**

4 The critical effects of kidney and liver pathology are systemic in nature. Therefore, EDC is
5 acting as a Category 3 gas for chronic exposure. For Category 3 gases, when available, animal
6 and human blood:gas partition coefficients are used to dosimetrically adjust for species
7 differences in toxicokinetics (TCEQ 2012).

8 $POD_{HEC} = POD_{ADJ} \times ((H_{b/g})_A / (H_{b/g})_H)$

9 where: $H_{b/g}$ = ratio of the blood:gas partition coefficient

10 A = animal

11 H = human

12 Because these data are lacking, a default value of 1 is used (TCEQ 2012).

13 $POD_{HEC} = POD_{ADJ} \times ((H_{b/g})_A / (H_{b/g})_H) = 2.0833 \text{ ppm} \times 1 = 2.0833 \text{ ppm}$

14 **4.1.6 Adjustments to the POD_{HEC}**

15 For the noncarcinogenic effects of EDC, UFs are applied to a POD to derive the chronic ReV
16 (i.e., assume a threshold MOA for a noncarcinogenic endpoint). The following UFs were
17 considered appropriate for application to the POD_{HEC} of 2.0833 ppm: 10 for UF_H , 3 for UF_A , and
18 6 for UF_D , for a total UF of 180.

- 19 • An UF_H of 10 was used to account for variation in sensitivity among the members of the
20 human population including possible child/adult differences, those with pre-existing medical
21 conditions, etc.;
- 22 • An animal-to-human UF_A of 3 was used to account for potential pharmacodynamic
23 differences between animals and humans (pharmacokinetic adjustment was already
24 performed); and
- 25 • A database deficiency UF_D of 6 was used because although there are several chronic studies
26 in multiple species available for EDC, including reproductive and developmental studies,
27 very few identified levels of toxicity below that which caused mortality. EDC shows a very
28 steep dose response curve (mild liver/kidney effects at 50 ppm and high toxicity/mortality at
29 250 ppm), which requires due consideration when selecting the UF_D given the studies
30 available. The quality of the study used as the POD is considered medium, and the
31 confidence in the acute database is medium to high.

32

$$\begin{aligned}
 1 \quad & \text{chronic ReV} = \text{POD}_{\text{HEC}} / (\text{UF}_{\text{H}} \times \text{UF}_{\text{A}} \times \text{UF}_{\text{D}}) \\
 2 \quad & = 2.0833 \text{ ppm} / (10 \times 3 \times 6) \\
 3 \quad & = 2.0833 \text{ ppm} / 180 \\
 4 \quad & = 0.011574 \text{ ppm} \\
 5 \quad & = 11.574 \text{ ppb or } 11 \text{ ppb (rounded to two significant digits)}
 \end{aligned}$$

6 **4.1.7 Health-Based Chronic ReV and ^{chronic}ESL_{threshold(nc)}**

7 In deriving the chronic ReV, no numbers were rounded between equations until the ReV was
8 calculated. The chronic ReV was rounded to two significant figures, resulting in a value of 11
9 ppb (44 µg/m³), and then used to calculate the ^{chronic}ESL_{threshold(nc)}. At the target hazard quotient of
10 0.3, the ^{chronic}ESL_{threshold(nc)} is 3.3 ppb (13 µg/m³) (Table 9).

11 **Table 9. Derivation of the Chronic ReV and ^{chronic}ESL**

Parameter	Values and Descriptions
Study	Spreafico et al. 1980
Study Population	Male and female SD rats and Swiss mice
Study Quality	Medium
Exposure Concentrations	Untreated, chamber control, 5, 10, 50, 150-250 ppm
Critical Effects	Increased ALT and uric acid in the serum, indicative of liver and kidney toxicity
POD (NOAEL)	10 ppm
Exposure Duration	7 h/d, 5 d/wk for 12 months
Extrapolation to continuous exposure (POD _{ADJ})	2.0833 ppm
POD _{HEC}	2.0833 ppm
Total UF	180
<i>Intraspecies UF</i>	10
<i>Interspecies UF</i>	3
<i>Incomplete Database UF</i>	6
<i>Database Quality</i>	Medium to high
Chronic ReV (HQ = 1)	44 µg/m³ (11 ppb)
^{chronic}ESL_{threshold(nc)} (HQ = 0.3)	13 µg/m³ (3.3 ppb)

1 **4.2 Carcinogenic Potential**

2 There has been some debate on the carcinogenic potential of EDC due to the varying results from
3 experimental tests, inconclusive data from epidemiological studies, and differences stemming
4 from route of exposure. Available data on the carcinogenicity of EDC are detailed below.

5 **4.2.1 Carcinogenic Weight of Evidence (WOE)**

6 EDC has been evaluated for carcinogenic potential by the International Agency for Research on
7 Cancer (IARC), the National Toxicology Program (NTP), USEPA, and the American
8 Conference of Industrial Hygienists (ACGIH) (Table 10). Generally, the TCEQ only performs
9 carcinogenic dose-response assessments for chemicals considered by the TCEQ either to be
10 “Carcinogenic to Humans” or “Likely to Be Carcinogenic to Humans” and for which available
11 data adequately characterize the dose-response curve.

12 **Table 10. Carcinogenic Weight of Evidence**

Group	Classification
IARC 1979, updated 1999	Possibly carcinogenic to humans
NTP 2011	Reasonably anticipated to be a human carcinogen
USEPA 1991	Probable human carcinogen
ACGIH 2001	Not classifiable as a human carcinogen

13 **4.2.2 Relevant Data**

14 **4.2.2.1 Epidemiological Studies**

15 Several studies have examined the correlation between excess cancer risk and EDC exposure.
16 However, these studies looked at industrial workers exposed to a number of chemicals including
17 EDC, so causality and significance are difficult to tease apart. Several of these studies are
18 detailed in IARC (1999) and ATSDR (2001) and a few are summarized here.

- 19
- 20 • Austin and Schnatter (1983) conducted a cohort study of 6588 petrochemical workers in
21 the U.S. that had reported an increased risk of brain cancers. There were 765 deaths and
22 150 deaths linked to cancer. The authors found an increased incidence in brain cancers,
23 but a nested study with the same group found no significant association between the risk
24 of primary brain tumors and exposure to EDC at the facility.
 - 25 • Benson and Teta (1993) examined mortality in 278 chlorohydrin production workers who
26 were exposed to EDC among other chemicals between 1940 and 1967. Out of 147 deaths,
27 40 were attributed to cancer, and increased incidences were observed in pancreatic,
lymphatic, and hematopoietic cancers. Since the workers were exposed to multiple

1 chemicals, including EDC, ethylene chlorohydrin, and bis(2-chloroethyl) ether, the
2 researchers were unable to link the cancer incidence with a particular chemical.

- 3 • Olsen et al. (1997) examined mortality in 1361 workers at two chlorohydrin production
4 plants similar to Benson and Teta (1993). There were a total of 300 deaths observed and
5 75 cancer deaths. The incidences of pancreatic, lymphatic, and hematopoietic cancers
6 were less than observed in Benson and Teta (1993), and no other correlations were
7 observed. This study lacked information on exposure concentrations and effects of
8 multiple chemical exposures, so no conclusions were made.

9 **4.2.2.2 Animal studies**

10 **4.2.2.2.1 Inhalation Studies**

11 Although several inhalation studies examining chronic exposure of EDC in animals have been
12 conducted, only a single study (Nagano et al. 2006) showed a statistically significant increase in
13 neoplastic lesions. IARC (1999) included a discussion of the same data that were originally
14 published in a study that examined several toxicants. The EDC-specific data were later published
15 in 2006 with a more detailed comparison to historical experimental tumor rates.

- 16 • Nagano et al. (2006) exposed male and female specific pathogen free F344/DuCrj rats and
17 Crj:BDF1 mice to EDC via inhalation at concentrations of 0, 10, 40, and 160 ppm (rats) and
18 0, 10, 30, and 90 ppm (mice) for 2 yr (50/sex/exposure group). These concentrations were
19 determined from an initial 13 wk study where rats and mice showed high mortality rates
20 when exposed to 320 and 160 ppm, respectively. No overt toxicity was observed at 160 ppm
21 in rats and a 9% and 7% decrease in body weight was observed in male and female mice,
22 respectively, at 80 ppm. EDC was vaporized by bubbling clean air through the liquid in a
23 temperature-regulated glass flask, and exposures were conducted in a 7600 L chamber for
24 rats and a 3700 L chamber for mice. Chamber concentrations were measured every 15 min,
25 with 2 yr averages of 10, 39.8, and 159.7 ppm for the rats, and 10, 30, and 89.8 ppm for the
26 mice. A number of non-neoplastic endpoints were examined and are discussed in Section
27 4.1.3.2.2. The incidences of the observed neoplastic lesions are presented in Tables 11 and 12
28 for rats and mice, respectively. In the rat study, there was a statistically significant increase in
29 the incidence of mammary gland fibroadenomas in males and subcutis fibromas, mammary
30 gland adenomas and mammary gland fibroadenomas in females following exposure to 160
31 ppm EDC compared to controls. There were also significant increasing trends in most of the
32 identified tumors, although the individual incidence may have not reached significance
33 except for one or two exposure groups at the most. The authors point out that the combined
34 incidence of mammary gland adenomas and fibroadenomas in female rats in the 40 ppm
35 exposure group is higher than the maximum tumor number identified in a historical records
36 comparison, however it did not reach statistical significance in the study. In the mouse study,
37 there was a statistically significant increase in the incidence of liver hemangiosarcomas in
38 males at 30 and 90 ppm and in malignant lymphomas in females at 10 and 30 ppm; however
39 neither showed a dose-response relationship nor a significant increasing trend. The observed

1 liver, lung, mammary gland, and uterine tumors in female mice did show a significant
2 increasing trend, however the individual incidence data did not reach significance.

3 **Table 11. Tumor incidence in rats exposed via inhalation to EDC**

Tumor Type/Incidence	Control	10 ppm	40 ppm	160 ppm
Males (#)	(50)	(50)	(50)	(50)
Subcutis fibroma ^a	6	9	12	15
Mammary gland adenoma	1	2	0	2
Mammary gland fibroadenoma ^a	0	0	1	5*
Combined mammary gland tumors ^a	1	2	1	7*
Peritoneum mesothelioma ^a	1	1	1	5
Females (#)	(50)	(50)	(50)	(50)
Subcutis fibroma ^a	0	0	1	5*
Mammary gland adenoma ^a	3	5	5	11*
Mammary gland fibroadenoma ^a	4	1	6	13*
Mammary gland adenocarcinoma ^a	1	2	0	5
Combined mammary gland tumors ^a	8	8	11	25*

4 * Significantly different from control at $p \leq 0.05$ by Fisher's exact test

5 ^a Significantly increasing trend at $p \leq 0.05$ by Peto's test

6

1 **Table 12. Tumor incidence in mice exposed via inhalation to EDC**

Tumor Type/Incidence	Control	10 ppm	30 ppm	90 ppm
Males (#)	(50)	(49)	(50)	(50)
Liver Hemangiosarcoma	0	4	6*	5*
Females (#)	(49)	(50)	(50)	(50)
Bronchiolo-alveolar adenoma ^a	4	1	3	8
Bronchiolo-alveolar carcinoma ^a	1	0	1	3
Combined bronchiolo-alveolar tumors ^a	5	1	4	11
Uterine endometrial stromal polyp ^a	2	0	1	6
Mammary Gland Adenocarcinoma ^a	1	2	1	6
Hepatocellular adenoma ^a	1	1	1	6
Hepatocellular carcinoma	1	0	1	0
Combined hepatocellular tumors ^a	2	1	2	6
Malignant lymphoma	6	17*	22*	12

2 * Significantly different from control at $p \leq 0.05$ by Fisher's exact test

3 ^a Significantly increasing trend at $p \leq 0.05$ by Peto's test

4 Several other inhalation studies showed either negative or mixed results.

- 5 • Cheever et al. (1990) exposed groups of 50 SD [CrI:CD(SD)BR outbred] rats to 50 ppm EDC
6 for 7 h/d, 5 d/wk for 2 yr (except for holidays). Animals were exposed in 2.2 m³ stainless
7 steel and glass chambers. EDC was volatilized by passing compressed air through liquid
8 EDC. Nominal concentrations were calculated on a daily basis, and analytical concentrations
9 were measured hourly. The average chamber concentration was 50.4 ppm for the target
10 concentration of 50 ppm. Body weights, food and water consumption were measured weekly.
11 Animals that either died during the study or were sacrificed after 2 yr were examined for
12 gross histopathological changes and neoplastic lesions. Although some tissue lesions and
13 masses were observed at an increased frequency, no statistically significant increase in any
14 neoplastic lesion was observed following 50 ppm EDC exposure. These results are consistent
15 with the lack of findings in rats chronically exposed to 40 ppm in Nagano et al. (2006).
16 • Spreafico et al. (1980) and Maltoni et al. (1980), as detailed in USEPA (2010) and ATSDR
17 (2001), exposed male and female SD rats and Swiss mice (90/sex/group) to inhalation of 5,
18 10, 50, or 150-250 ppm EDC for 7 h/d, 5 d/wk, for up to 18 months. Spreafico et al. (1980)
19 reported on the methods and chronic toxicity of EDC, while Maltoni et al. (1980) reported
20 the neoplastic endpoints. Control groups consisted of 180 rats (chamber control and

1 untreated) and 249 mice (untreated only). Originally the highest exposure concentration was
2 set at 250 ppm, but due to high toxicity in the animals after several days, the concentration
3 was reduced to 150 ppm. Body weights were recorded every 2 wks, and 8-10 animals from
4 each group were sacrificed at 3, 6, and 18 months. No differences were noted in the tumor
5 incidence compared to controls in mice; however, a detailed report was not provided.
6 Although the USEPA (2010) reported no differences in tumor incidence in rats compared to
7 controls, IARC (1979) reported the incidence of mammary fibromas and fibroadenomas to be
8 statistically significantly increased in the 5 ppm (56/90), 50 ppm (49/90), and 150-250 ppm
9 groups (47/90) compared to chamber controls (26/90), but not in the 10 ppm group (33/90).
10 However, they were not significantly different from the untreated controls (47/90), the two
11 control groups were significantly different from each other, and the increased incidence did
12 not show a dose-response (e.g., there was actually a 10% decrease in incidence between 5
13 and 150-250 ppm). Notwithstanding the lack of a dose-response, the finding of potential
14 increases in mammary fibromas and fibroadenomas at 150-250 ppm is consistent with the
15 statistically elevated incidences for these endpoints found in female rats exposed to 160 ppm
16 in Nagano et al. (2006).

17 **4.2.2.2.2 Oral Study**

18 The USEPA (1991) used an oral gavage study of chronic EDC exposure to derive its inhalation
19 unit risk factor (URF) through route-to-route extrapolation.

- 20 • The National Cancer Institute (NCI 1978) exposed Osborne-Mendel rats and B6C3F1 mice
21 (50/species/sex/group) to EDC by oral gavage alongside untreated and vehicle-treated
22 controls (20/species/sex/group). Animals were treated 5 d/wk for 78 wks with time-weighted
23 average doses of 47 and 95 mg/kg/d for rats, 97 and 195 mg/kg/d for male mice, and 149 and
24 299 mg/kg/d for female mice. Since the doses varied each week, the authors represented the
25 time-weighted average doses as the sum of the doses divided by the total weeks of exposure.
26 For the rat study, mortality was high in the group exposed to 95 mg/kg/d. An increased
27 incidence of squamous-cell carcinomas was observed in the forestomachs of male rats in the
28 95 mg/kg/d group (9/50) and the 47 mg/kg/d group (3/50) compared to controls (0/40). The
29 incidence of hemangiosarcomas was also increased in the male rats in the 95 mg/kg/d group
30 (7/27) and the 47 mg/kg/d group (9/48) compared to controls (0/40). Female rats showed a
31 significant increase in mammary adenocarcinomas. In the mouse study, female mice showed
32 a statistically significant increase in mortality as the dose increased, while male rats showed
33 no association. Mice showed increased incidences of hepatocellular carcinomas and
34 alveolar/bronchiolar adenomas.

35 The USEPA (1991) used the increased incidence of hemangiosarcomas as a POD for developing
36 an inhalation URF. The human equivalent dose was calculated assuming an average human
37 weight of 70 kg and an average rat weight of 0.5 kg. The 95% upper bound of risk was
38 calculated using 90 weeks of exposure. The calculated USEPA inhalation URF was 2.6E-05 per

1 $\mu\text{g}/\text{m}^3$ (1.05E-04 per ppb), which corresponds to an air concentration of $0.4 \mu\text{g}/\text{m}^3$ at an excess
2 risk level of 1E-05 (1 in 100,000) (i.e., $1\text{E}-05 / 2.6\text{E}-05 \text{ per } \mu\text{g}/\text{m}^3 = 0.4 \mu\text{g}/\text{m}^3$).

3 **4.2.2.2.1 Comparison Studies**

4 A few studies have examined the possible differences in toxicity following exposure to EDC
5 through various routes. ATSDR (2001) states that inhalation exposure to EDC is predicted to
6 produce less metabolites in the liver and lungs than equivalent oral exposures, which may
7 explain some of the differences observed between routes of exposure.

- 8 • Storer et al. (1984) compared the hepatotoxicity and genotoxicity following EDC exposure of
9 male C57BL/6 x C3HF₁ mice by inhalation, oral gavage, and intraperitoneal (i.p.) injection.
10 For the inhalation route, exposures were performed in a 30L stainless steel and glass
11 chamber, and EDC was vaporized in a gas washing bottle by a metered air flow.
12 Concentrations were monitored at 15-min intervals, with target concentrations of 150, 500,
13 1000, and 2000 ppm (time-weighted average concentrations of 158, 499, 1072, and 1946
14 ppm). For the gavage and i.p. routes, animals were treated with 200, 300, 400, 500, and 600
15 mg/kg EDC. All animals were exposed for 4 h and sacrificed 24 h later. The researchers
16 collected tissues and blood samples to measure hepatotoxicity and nephrotoxicity, and
17 assessed DNA damage in the liver by measuring the amount of double-stranded DNA that
18 could be recovered (inversely proportional to DNA damage from double-stranded breaks).
19 No evidence of DNA damage in the liver was observed at 150 or 500 ppm following a 4 h
20 inhalation exposure. At the higher exposures of 1000 and 2000 ppm, some DNA damage was
21 observed; however, there was also a high level of exposure-related mortality. DNA damage
22 was detected at lower doses following i.p. and gavage exposure. The authors concluded that
23 EDC is genotoxic at non-necrogenic dose levels following i.p. or gavage exposures, but non-
24 genotoxic at comparable doses following inhalation exposure.
- 25 • Hooper et al. (1980) compared the data from the oral exposure study by NCI (1978) where
26 several types of tumors were observed and the inhalation exposure study by Spreafico et al.
27 (1980) and Maltoni et al. (1980) where there was no significant carcinogenic effect. Hooper
28 et al. examined several different variables that could have played a role in the observed
29 differences, including the purity of the chemical, dose levels, routes of exposure, strain
30 differences, and statistical significance. The chemical purity appeared to be similar in the two
31 studies, although the study by NCI (1978) tested several different chemicals at the same time,
32 so other contaminants in the air may have been responsible for the observed effects. When
33 conducting route-to-route calculations, it was determined that the inhalation study used doses
34 that were much lower than the oral study, although the two highest doses (50 and 150 ppm)
35 in rats (calculated to be 16 and 48 mg/kg/d) and mice (calculated to be 56 and 171 mg/kg/d)
36 were comparable to the doses used in the oral gavage study (rats exposed to 24 and 48
37 mg/kg/d, male mice exposed to 60 and 120 mg/kg/d, and female mice exposed to 92.5 and
38 185 mg/kg/d). For differences in the route of exposure, the authors state that although the
39 blood concentration, metabolism, and tissue distribution of EDC are approximately the same
40 for oral and inhalation routes of exposure, the possibility of gut flora producing carcinogenic

1 metabolites cannot be ruled out. Hooper et al. (1980) also noted that there was a high level of
2 early mortality in both studies, which could reduce the number of observed tumors as they
3 tend to form later in life. They concluded that the observed difference in the carcinogenic
4 potential of EDC in these two studies was most likely due to multiple factors, but that further
5 data including a life-table analysis would be required to tease out the specific cause.

6 **4.2.3 Carcinogenic MOA**

7 Absorption, distribution, and metabolism of EDC following inhalation or ingestion are rapid and
8 complete in rats, with 85% of the metabolites being excreted in urine (OEHHA 2000).
9 Metabolism can occur through two different pathways: cytochrome P450 and glutathione S-
10 transferase (IARC 1997). Animal studies suggest that EDC may be metabolized to 2-
11 chloroacetaldehyde and other reactive metabolites, which are able to bind and inhibit cellular
12 macromolecules (ASTDR 2001). Most metabolites of EDC are conjugated with glutathione and
13 are considered nontoxic with the exception of S-(2-chloroethyl)glutathione, which has been
14 shown to form DNA adducts (IARC 1997). However, the metabolism of EDC in humans has not
15 been studied, and the specific MOA for the potential EDC carcinogenicity remains unknown.
16 Therefore, a non-threshold MOA was assumed for developing a ^{chronic}ESL_{nonthreshold(c)}.

17 **4.2.4 Genotoxicity**

18 A detailed summary of the genotoxic effects of EDC can be found in ATSDR (2001) and IARC
19 (1999). No studies were available on the genotoxicity of EDC in humans following inhalation or
20 oral exposure. Storer et al. (1984) reported an increase in double-stranded breaks in mice
21 following a 4-h exposure to 1000 ppm EDC, although this concentration also induced significant
22 mortality. Single-stranded breaks were also observed in rat liver following oral gavage of EDC
23 (IARC 1999). *In vitro*, EDC has been shown in numerous studies to interact with DNA and
24 induce genotoxic effects (ASTDR 2001). EDC was mutagenic in *Salmonella typhimurium* and
25 *Drosophila melanogaster*, as well as mouse and rat liver, lung, and kidney cells (IARC 1999). A
26 detailed list of the positive genotoxic assays and their results can be found in ASTDR (2001).

27 **4.2.5 POD for Key Study and Critical Effect**

28 The Nagano et al. (2006) study will be used as the key study as it is the inhalation study that
29 most clearly showed a statistically significant increase for certain tumor types, some of which
30 also showed a statistically significant increasing trend across exposure groups and a dose-
31 response. Liver hemangiosarcomas in mice showed a significant increase at 30 ppm, and had a
32 close dose-response relationship (0/50 in controls, 4/49 at 10 ppm, 6/50 at 30 ppm, 5/50 at 90
33 ppm), so this response was chosen as a candidate cancer endpoint. USEPA (1991) used the same
34 cancer endpoint in male rats to derive their URF, although based on a different route of exposure
35 (i.e., the oral exposure study by NCI 1978), which strengthens the use of this candidate cancer
36 endpoint. Combined mammary tumors in rats showed a dose-response relationship, but incidence
37 was only significant at the highest dose tested (160 ppm), five times higher than the
38 concentration that produced significant increases in liver hemangiosarcomas in mice. Increased

1 incidence of mammary tumors at the highest concentration was also observed in the oral study
2 by NCI (1978), and the USEPA chose liver hemangiosarcomas as their critical cancer endpoint.
3 Toxicity studies confirm that the liver is a target organ of EDC, further supporting the use of
4 liver hemangiosarcomas as the cancer endpoint. Malignant lymphomas in mice were also
5 considered, but the data showed a clearly non-monotonic dose-response (e.g., incidences of
6 17/49 at 10 ppm and 22/50 at 30 ppm, with a much lower incidence of 12/50 at 90 ppm).

7 The TCEQ performed Benchmark Concentration (BMC) modeling using USEPA Benchmark
8 Dose (BMD) software (version 2.6) for the data in Table 12 which was taken from the Nagano et
9 al. (2006) study. Data were used to predict 95% lower confidence limits on the BMCs using
10 dichotomous models. A default benchmark response (BMR) of 10% was selected for extra risk
11 (BMC_{10}) and $BMCL_{10}$. All of the available dichotomous and multistage cancer models were run
12 with (Appendices 1.1 and 1.2) and without (Appendices 1.3 and 1.4) including the highest dose,
13 and the results can be found in Appendix 1. The best fit model based in the lowest $BMCL_{10}$ and
14 the best fit to the curve is listed in Table 13 along with the detailed results.

15 **Table 13. BMC Results for the Best Fit Model**

Endpoint	Best Model	p value	AIC	Scaled Residual	BMC_{10}	$BMCL_{10}$
Liver Hemangiosarcomas	Dichotomous-Hill model using all data points	0.749	103.01	0	11.1 ppm	9.02 ppm

16 Using the incidence of liver hemangiosarcomas from the Nagano et al. (2006) study, a BMC_{10} or
17 effective concentration (EC_{10}) of 11.1 ppm was calculated, giving a $BMCL_{10}$ or lower EC
18 (LEC_{10}) of 9.02 ppm (36.51 mg/m^3). These values are very close together, strengthening the use
19 of this model. From this data, an inhalation URF can be derived using the following equation
20 (TCEQ 2012):

21
$$URF = 0.1 / BMCL_{10}$$

22
$$URF = 0.1 / 9.02 \text{ ppm} = 0.0111 \text{ (ppm)}^{-1} \text{ or } 0.002739 \text{ (mg/m}^3\text{)}^{-1}$$

23
$$URF = 1.1E-05 \text{ (ppb)}^{-1} \text{ or } 2.7E-06 \text{ (}\mu\text{g/m}^3\text{)}^{-1} \text{ (rounded to two significant figures)}$$

24 The central estimate using the BMC_{10} to calculate the URF is $9.0E-06 \text{ (ppb)}^{-1}$.

25 **4.2.6 Calculation of an Air Concentration at 1×10^{-5} Excess Cancer Risk**

26 The calculated URF based on increased incidence of liver hemangiosarcomas from the Nagano
27 et al. (2006) study is $1.1E-05 \text{ (ppb)}^{-1}$ or $2.7E-06 \text{ (mg/m}^3\text{)}^{-1}$. The no significant risk level of $1E-05$
28 is calculated as follows (TCEQ 2012):

29
$${}^{\text{chronic}}ESL_{\text{nonthreshold(c)}} = 1E-05 / URF$$

1 $= 1\text{E-}05 / 1.1\text{E-}05 \text{ (ppb)}^{-1}$
2 $= 0.91 \text{ ppb (} 3.7 \text{ }\mu\text{g/m}^3\text{)}$

3 **4.2.7 Comparison of Cancer Potency Factors**

4 Table 14 lists the inhalation URF and toxicity values calculated at a cancer risk level of 1E-05
5 that are available. Both the USEPA and OEHHA toxicity values are based on route-to-route
6 extrapolation from oral studies.

7 **Table 14. Available Inhalation URFs and Chronic Toxicity Value**

Agency	Inhalation URF	Chronic Toxicity Value
TCEQ ^{chronic} ESL _{nonthreshold(c)}	2.7E-06 ($\mu\text{g/m}^3$) ⁻¹	3.7 $\mu\text{g/m}^3$
OEHHA (2009)	2.1E-05 ($\mu\text{g/m}^3$) ⁻¹	0.5 $\mu\text{g/m}^3$
USEPA (1991)	2.6E-05 ($\mu\text{g/m}^3$) ⁻¹	0.4 $\mu\text{g/m}^3$

8 **4.2.8 Evaluating Susceptibility from Early-Life Exposures**

9 TCEQ (2012) states that carcinogens acting through a mutagenic MOA need to be evaluated for
10 the potential increase in cancer due to early-life exposures compared with adult and whole-life
11 exposure. USEPA (2005) provides default age-dependent adjustment factors (ADAFs) to
12 account for potential increased susceptibility in children due to early-life exposure when a
13 chemical has been identified as acting through a mutagenic MOA for carcinogenesis. Storer et al.
14 (1984) reported an increase in double-strand DNA breaks in mice following a 4-h exposure to
15 1000 ppm EDC (although this concentration also induced significant mortality), and single-
16 strand breaks were also observed in rat liver following oral gavage of EDC (IARC 1999). *In*
17 *vitro*, EDC has been shown to interact with DNA and induce genotoxic effects (ASTDR 2001).
18 EDC was mutagenic in *Salmonella typhimurium* and *Drosophila melanogaster*, as well as mouse
19 and rat liver, lung, and kidney cells (IARC 1999), but it did not induce micronuclei in mouse
20 cells (ATSDR 2001). Although these changes suggest the potential for genotoxic and/or
21 mutagenic effects, once a carcinogen has been determined to have mutagenic potential, there are
22 several important considerations in assessing evidence for a mutagenic MOA for cancer. For
23 example: (1) whether the chemical-induced mutation occurs prior to the initiation of the
24 carcinogenic process (i.e., early in relation to the key events that lead to cancer) in the target
25 tissue (i.e., site and temporal concordance between mutagenicity and carcinogenicity), and if so
26 (2) whether the chemical-induced mutation is the key event that initiates the carcinogenic
27 process in the target tissue. See Section 5.7.5.1.2 of TCEQ (2012) for additional information,
28 including a hierarchy for types of relevant evidence. Most importantly, for a chemical to act by a
29 mutagenic MOA, either the chemical or its direct metabolite must be the agent inducing the
30 mutations that initiate cancer in the target tissue. As there is no default carcinogenic MOA, the
31 scientific burden of proof is a reasonably robust demonstration through direct evidence that the
32 specific mutation(s) caused by the chemical or its metabolite is in fact the first step in target

1 tissue which initiates a cascade of other key events that are critical to the carcinogenic process in
2 the specific tumors. Mere plausibility (whether or not information on other possible MOAs is
3 available) based on the consistency of circumstantial evidence is not tantamount to an adequately
4 robust demonstration that mutagenicity is in fact the initiating event in target tissues. Thus, if the
5 weight of evidence supports a chemical's genotoxic and/or mutagenic potential, for evaluation of
6 the MOA emphasis should then be placed on evidence of the chemical's mutagenicity being the
7 critical, initiating carcinogenic event in target cells (at relevant doses if possible). In the event
8 scientifically convincing data on the carcinogenic MOA are lacking, the carcinogenic MOA may
9 ultimately be judged simply to be unknown or not sufficiently elucidated or established (TCEQ
10 2012). This is the case for EDC, for which the carcinogenic MOA is certainly unclear. As the
11 MOA for EDC has not been demonstrated to be mutagenic, consistent with TCEQ guidance
12 (TCEQ 2012), ADAFs will not be applied to the URF at this time. This issue will be reevaluated
13 periodically as new scientific information on EDC's carcinogenic MOA becomes available.

14 ***4.3 Welfare-Based Chronic ESL***

15 No data were found regarding long-term vegetation effects.

16 ***4.4 Long-Term ESL and Values for Air Monitoring Evaluation***

17 The chronic evaluation resulted in the derivation of the following values:

- 18 • Chronic ReV = $44 \mu\text{g}/\text{m}^3$ (11 ppb)
- 19 • $\text{chronicESL}_{\text{threshold(nc)}} = 13 \mu\text{g}/\text{m}^3$ (3.3 ppb)
- 20 • URF = $2.7\text{E-}06 (\mu\text{g}/\text{m}^3)^{-1}$ ($1.1\text{E-}05 (\text{ppb})^{-1}$)
- 21 • $\text{chronicESL}_{\text{nonthreshold(c)}} = 3.7 \mu\text{g}/\text{m}^3$ (0.91 ppb)

22 The long-term ESL for air permit reviews is the $\text{chronicESL}_{\text{nonthreshold(c)}}$ of $3.7 \mu\text{g}/\text{m}^3$ (0.91 ppb).
23 The chronic ReV of $44 \mu\text{g}/\text{m}^3$ (11 ppb) is for the evaluation of ambient air monitoring data. The
24 $\text{chronicESL}_{\text{threshold(nc)}}$ (HQ = 0.3) would not be used to evaluate ambient air monitoring data (Table
25 2).

26 ***4.5 Noncarcinogenic Chronic Inhalation Observed Adverse Effect Level***

27 Observed inhalation adverse effect levels are described in more detail in Section 3.4 and in
28 TCEQ 2012. This section is for noncarcinogenic effects only; variability in the carcinogenic data
29 make it unsuitable for determination of an observed adverse effect level. The chronic POD of 10
30 ppm determined from the Spreafico et al. (1980) and Maltoni et al. (1980) study was based on
31 liver and kidney effects observed in rats following 50 ppm EDC inhalation exposure. The
32 LOAEL of 50 ppm, where effects occurred in some animals, represents a concentration at which
33 similar effects could possibly occur in some individuals exposed over the same duration or
34 longer. Based on the TCEQ guidelines (2012), no duration adjustment is needed; however an
35 animal-to-human dosimetric adjustment is used to calculate the $\text{LOAEL}_{\text{HEC}}$. Since the RGDR is
36 1, based on updated guidelines from the USEPA (2012), the $\text{LOAEL}_{\text{HEC}}$ is equal to the LOAEL

1 of 50 ppm. Effects are not a certainty as there may be inter- and intraspecies differences in
2 sensitivity. The chronic inhalation observed adverse effect level of 50 ppm is provided for
3 informational purposes only (TCEQ 2012). As the basis for development of inhalation observed
4 adverse effect levels is limited to available data, future studies could possibly identify a lower
5 POD for this purpose.

6 The margin of exposure between the observed adverse effect level (50 ppm) and the chronic ReV
7 (0.011 ppm) is a factor of approximately 4,500.

8 **Chapter 5 References**

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1 **Appendix 1 Benchmark Concentration (BMC) Modeling**

2 The TCEQ performed Benchmark Concentration (BMC) modeling using USEPA Benchmark
3 Dose (BMD) software (version 2.6) for the incidence of liver hemangiosarcomas presented in
4 Table 12 which was taken from the Nagano et al. (2006). Data were used to predict 95% lower
5 confidence limits on the BMCs using dichotomous models. A default BMR of 10% was selected
6 for extra risk (BMC_{10}) and $BMCL_{10}$. All of the available dichotomous and multistage cancer
7 models were run with (Appendices 1.1 and 1.2) and without (Appendices 1.3 and 1.4) including
8 the highest dose. All of the models are presented below, with the best fit model based in the
9 lowest $BMCL_{10}$ and the best fit to the curve shown in bold and graphically below its respective
10 table.

1 **Appendix 1.1 Dichotomous models using all of the doses**

Tumor Type/Incidence	Control	10 ppm	30 ppm	90 ppm
Males (#)	(50)	(49)	(50)	(50)
Liver Hemangiosarcoma	0	4	6*	5*

2 **Table 15. Dichotomous models using all of the data points for liver hemangiosarcomas**

Model ^a	Goodness of fit		BMD _{10Pct}	BMDL _{10Pct}	Basis for model selection
	p-value	AIC			
Gamma ^b	0.0694	107.76	84.0	36.4	Of the models that provided an adequate fit and a valid BMDL estimate, the Dichotomous-Hill model was selected based on lowest BMDL.
Dichotomous-Hill	0.749	103.01	11.1	9.02	
Logistic	0.0682	108.60	116	65.8	
LogLogistic	0.0694	107.61	77.5	31.0	
Probit	0.0687	108.52	114	62.8	
LogProbit	0.852	101.22	28.6	error ^c	
Weibull ^d	0.0694	107.76	84.0	36.4	
Quantal-Linear ^e					
Multistage 4 ^o	error	error	error ^c	error ^c	
Multistage 3 ^{of}	0.0694	107.76	84.0	36.4	
Multistage 2 ^{og}					

3 ^a Selected model in bold; scaled residuals for selected model for doses 0, 10, 30, and 90 were 0, 0, 0.23, -
4 0.23, respectively.

5 ^b The Gamma model may appear equivalent to the Weibull model, however differences exist in digits not
6 displayed in the table. This also applies to the Multistage 3^o model. This also applies to the Multistage 2^o
7 model. This also applies to the Quantal-Linear model.

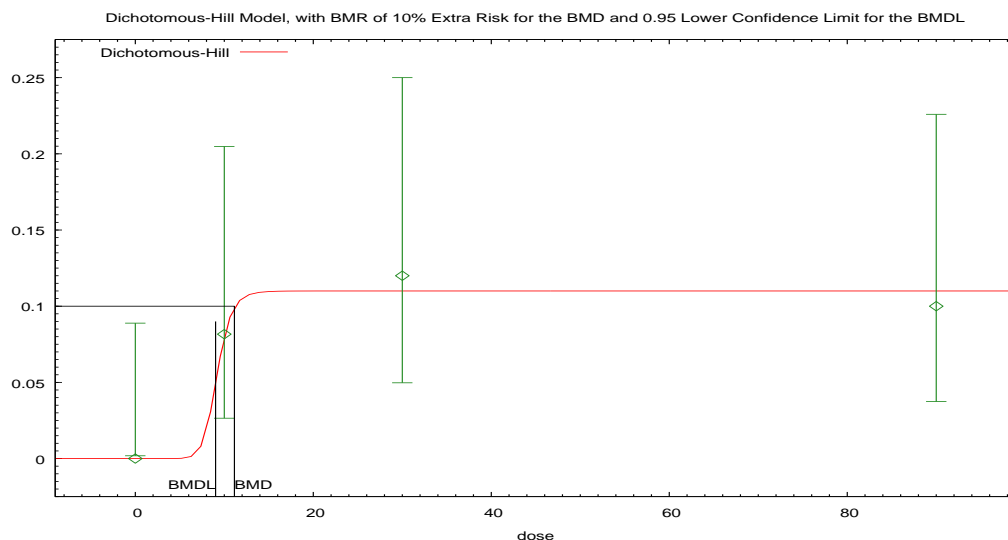
8 ^c BMD or BMDL computation failed for this model.

9 ^d For the Weibull model, the power parameter estimate was 1. The models in this row reduced to the
10 Quantal-Linear model.

11 ^e The Quantal-Linear model may appear equivalent to the Gamma model, however differences exist in
12 digits not displayed in the table. This also applies to the Multistage 3^o model. This also applies to the
13 Multistage 2^o model.

14 ^f For the Multistage 3^o model, the beta coefficient estimates were 0 (boundary of parameters space). The
15 models in this row reduced to the Multistage 2^o model.

16 ^g The Multistage 2^o model may appear equivalent to the Gamma model, however differences exist in
17 digits not displayed in the table. This also applies to the Weibull model. This also applies to the Quantal-
18 Linear model.



1 10:48 03/04 2015

2 **Figure 1. Plot of incidence rate by dose with fitted curve for Dichotomous-Hill model; dose**
3 **shown in ppm.**

4 **Dichotomous Hill Model.** (Version: 1.3; Date: 02/28/2013)

5 The form of the probability function is: $P[\text{response}] = v * g + (v - v * g) / [1 + \text{EXP}(-\text{intercept} - \text{slope} * \text{Log}(\text{dose}))]$

6
7 Slope parameter is restricted as slope ≥ 1

8 **Benchmark Dose Computation.**

9 BMR = 10% Extra risk

10 BMD = 11.1405

11 BMDL at the 95% confidence level = 9.02383

12 **Parameter Estimates**

Variable	Estimate	Default Initial Parameter Values
v	0.11	1
g	0	0
intercept	-2.5498E+01	-5.9688E+00
slope	11.5328	1

13 **Analysis of Deviance Table**

Model	Log(likelihood)	# Param's	Deviance	Test d.f.	p-value
Full model	-48.45	4			

Fitted model	-48.51	3	0.102272	1	0.75
Reduced model	-53.2	1	9.48825	3	0.02

1 AIC: = 103.011

2 **Goodness of Fit Table**

Dose	Est. Prob.	Expected	Observed	Size	Scaled Resid
0	0	0	0	50	0
10	0.0816	4	4	49	0
30	0.11	5.5	6	50	0.23
90	0.11	5.5	5	50	-0.23

3 $\text{Chi}^2 = 0.1$

4 d.f = 1

5 P-value = 0.7493

1 **Appendix 1.2 Multistage cancer models using all of the doses**

Tumor Type/Incidence	Control	10 ppm	30 ppm	90 ppm
Males (#)	(50)	(49)	(50)	(50)
Liver Hemangiosarcoma	0	4	6*	5*

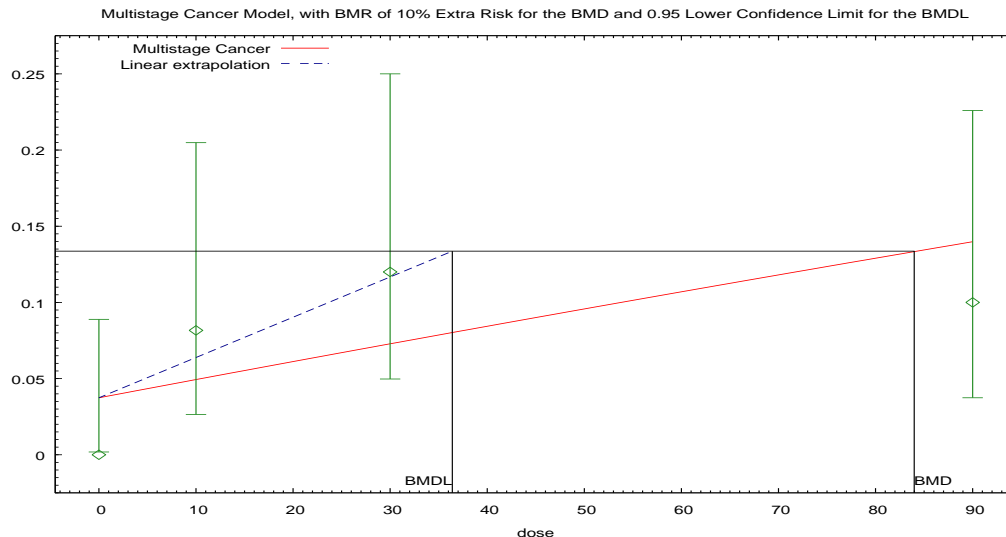
2

3 **Table 16. Multistage cancer models using all of the data points for liver hemangiosarcomas**

Model ^a	Goodness of fit		BMD _{10Pct}	BMDL _{10Pct}	Basis for model selection
	p-value	AIC			
Four	error	error	error ^b	error ^b	Of the models that provided an adequate fit and a valid BMDL estimate, the Multistage 1 model was selected based on the simplest model.
Three	0.0694	107.76	84.0	36.4	
Two					
One	0.0694	107.76	84.0	36.4	

4 ^a Selected model in bold; scaled residuals for selected model for doses 0, 10, 30, and 90 were -1.39, 1.04,
5 1.28, -0.82, respectively.

6 ^b BMD or BMDL computation failed for this model.



7

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8 **Figure 2. Plot of incidence rate by dose with fitted curve for Multistage-Cancer 1° model;**
9 **dose shown in ppm.**

10 **Multistage Model.** (Version: 3.4; Date: 05/02/2014)

11 The form of the probability function is: $P[\text{response}] = \text{background} + (1-\text{background}) * [1-\text{EXP}(-$
12 $\text{beta1} * \text{dose}^1 - \text{beta2} * \text{dose}^2 \dots)]$

- 1 The parameter betas are restricted to be positive
- 2 **Benchmark Dose Computation.**
- 3 BMR = 10% Extra risk
- 4 BMD = 83.982
- 5 BMDL at the 95% confidence level = 36.4061
- 6 BMDU at the 95% confidence level = 2.29572E+43
- 7 Taken together, (36.4061, 2.29572E+43) is a 90% two-sided confidence interval for the BMD
- 8 Multistage Cancer Slope Factor = 0.00274679

9 **Parameter Estimates**

Variable	Estimate	Default Initial Parameter Values
Background	0.0373594	0.0526664
Beta(1)	0.00125456	0.000784122

10 **Analysis of Deviance Table**

Model	Log(likelihood)	# Param's	Deviance	Test d.f.	p-value
Full model	-48.45	4			
Fitted model	-51.88	2	6.84668	2	0.03
Reduced model	-53.2	1	9.48825	3	0.02

11 AIC: = 107.756

12 **Goodness of Fit Table**

Dose	Est. Prob.	Expected	Observed	Size	Scaled Resid
0	0.0374	1.868	0	50	-1.39
10	0.0494	2.419	4	49	1.04
30	0.0729	3.646	6	50	1.28
90	0.1401	7.007	5	50	-0.82

13 Chi² = 5.34

14 d.f = 2

15 P-value = 0.0694

1 **Appendix 1.3 Dichotomous models dropping the highest dose**

Tumor Type/Incidence	Control	10 ppm	30 ppm
Males (#)	(50)	(49)	(50)
Liver Hemangiosarcoma	0	4	6*

2

3 **Table 17. Dichotomous models dropping the highest dose for liver hemangiosarcomas**

Model ^a	Goodness of fit		BMD _{10Pct}	BMDL _{10Pct}	Basis for model selection
	p-value	AIC			
Gamma ^b Weibull ^c Multistage 2° Quantal-Linear	0.552	67.480	19.8	12.3	Of the models that provided an adequate fit and a valid BMDL estimate, the LogLogistic model was selected based on lowest BMDL.
Dichotomous-Hill Multistage 4° Multistage 3°	error	error	error ^f	error ^f	
Logistic	0.0841	72.296	28.5	21.1	
LogLogistic	0.606	67.329	19.4	11.5	
Probit	0.0892	72.090	27.7	20.0	
LogProbit	1.000	68.401	17.6	error ^d	

4 ^a Selected model in bold; scaled residuals for selected model for doses 0, 10, and 30 were 0, 0.85, -0.54,
5 respectively.

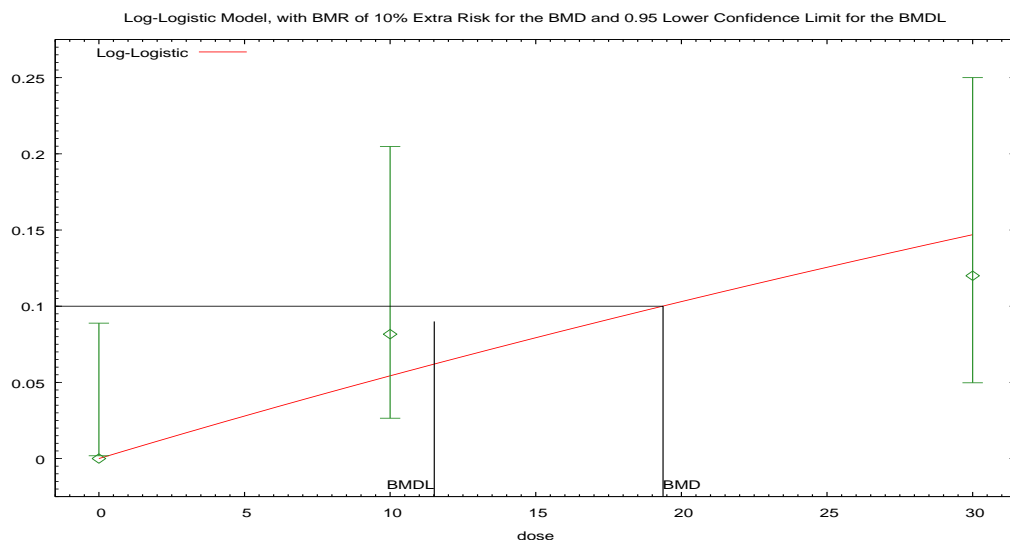
6 ^b For the Gamma and Weibull models, the power parameter estimates were 1 (boundary of parameter
7 space).For the Gamma model, the power parameter estimate was 1. The model is equivalent to the
8 Quantal-Linear model.

9 ^c For the Weibull and Gamma models, the power parameter estimates were 1 (boundary of parameter
10 space).For the Weibull model, the power parameter estimate was 1. The models in this row reduced to the
11 Quantal-Linear model.

12 ^d BMD or BMDL computation failed for this model.

13 ^e For the Multistage 4° model, the beta coefficient estimates were 0 (boundary of parameters space). The
14 models in this row reduced to the Multistage 3° model.

15 ^f BMD or BMDL computation failed for this model



1 09:09 03/04 2015

2 **Figure 3. Plot of incidence rate by dose with fitted curve for LogLogistic model; dose shown**
3 **in ppm.**

4 **Logistic Model.** (Version: 2.14; Date: 2/28/2013)

5 The form of the probability function is: $P[\text{response}] = \text{background} + (1 - \text{background}) / [1 + \text{EXP}(-$
6 $\text{intercept} - \text{slope} * \text{Log}(\text{dose}))]$

7 Slope parameter is restricted as slope ≥ 1

8 **Benchmark Dose Computation.**

9 BMR = 10% Extra risk

10 BMD = 19.3685

11 BMDL at the 95% confidence level = 11.5142

12 **Parameter Estimates**

Variable	Estimate	Default Initial Parameter Values
background	0	0
intercept	-5.1609E+00	-4.9705E+00
slope	1	1

13 **Analysis of Deviance Table**

Model	Log(likelihood)	# Param's	Deviance	Test d.f.	p-value
Full model	-32.2	3			
Fitted model	-32.66	1	0.927873	2	0.63

Reduced model	-36.67	1	8.93963	2	0.01
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1 AIC: = 67.3288

2 **Goodness of Fit Table**

Dose	Est. Prob.	Expected	Observed	Size	Scaled Resid
0	0	0	0	50	0
10	0.0543	2.658	4	49	0.85
30	0.1468	7.342	6	50	-0.54

3 $\chi^2 = 1$

4 d.f = 2

5 P-value = 0.6056

1 **Appendix 1.4 Multistage cancer models dropping the highest dose**

Tumor Type/Incidence	Control	10 ppm	30 ppm
Males (#)	(50)	(49)	(50)
Liver Hemangiosarcoma	0	4	6*

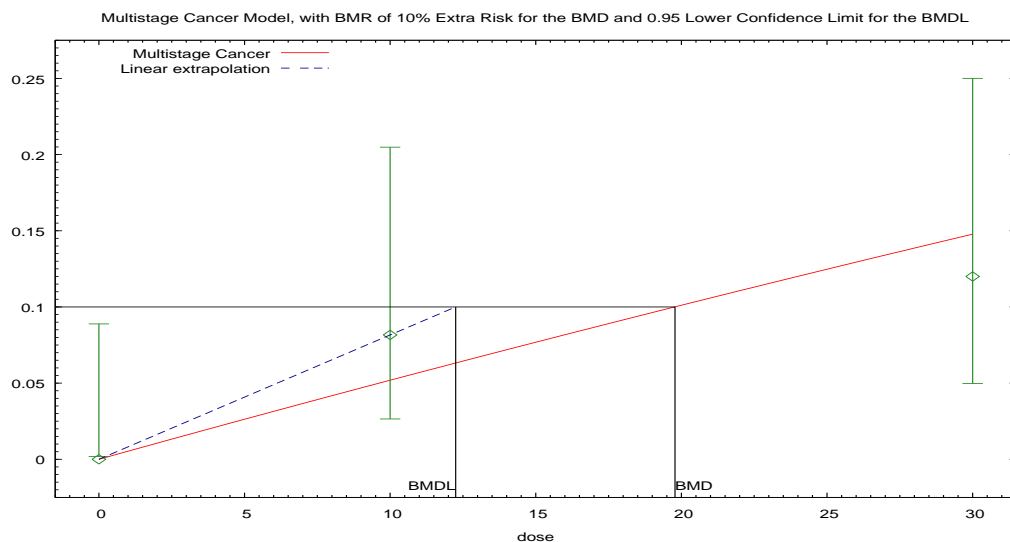
2 **Table 18. Multistage cancer models dropping the highest dose for liver hemangiosarcomas**

Model ^a	Goodness of fit		BMD _{10Pct}	BMDL _{10Pct}	Basis for model selection
	p-value	AIC			
Four Three	error	error	error ^c	error ^c	Of the models that provided an adequate fit and a valid BMDL estimate, the Multistage 1 model was selected based on the simplest model.
Two One	0.552	67.480	19.8	12.3	

3 ^a Selected model in bold; scaled residuals for selected model for doses 0, 10, and 30 were 0,
4 0.94, -0.55, respectively.

5 ^b BMD or BMDL computation failed for this model.

6 ^c BMD or BMDL computation failed for this model



7
8 **Figure 4. Plot of incidence rate by dose with fitted curve for Multistage-Cancer 1° model;**
9 **dose shown in ppm.**

10 **Multistage Model.** (Version: 3.4; Date: 05/02/2014)

11 The form of the probability function is: $P[\text{response}] = \text{background} + (1-\text{background}) * [1-\text{EXP}(-$
12 $\text{beta1} * \text{dose}^{\text{beta2}} * \text{dose}^{\text{beta1}} \dots)]$

13 The parameter betas are restricted to be positive

1 **Benchmark Dose Computation.**

2 BMR = 10% Extra risk

3 BMD = 19.7807

4 BMDL at the 95% confidence level = 12.2507

5 BMDU at the 95% confidence level = 42.1603

6 Taken together, (12.2507, 42.1603) is a 90% two-sided confidence interval for the BMD

7 Multistage Cancer Slope Factor = 0.00816282

8 **Parameter Estimates**

Variable	Estimate	Default Initial Parameter Values
Background	0	0.0180691
Beta(1)	0.00532644	0.00395721

9 **Analysis of Deviance Table**

Model	Log(likelihood)	# Param's	Deviance	Test d.f.	p-value
Full model	-32.2	3			
Fitted model	-32.74	1	1.07919	2	0.58
Reduced model	-36.67	1	8.93963	2	0.01

10 AIC: = 67.4801

11 **Goodness of Fit Table**

Dose	Est. Prob.	Expected	Observed	Size	Scaled Resid
0	0	0	0	50	0
10	0.0519	2.542	4	49	0.94
30	0.1477	7.384	6	50	-0.55

12 $\text{Chi}^2 = 1.19$

13 d.f = 2

14 P-value = 0.5524